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(54) Title:  $\alpha$ 1-3 GALACTOSYLTRANSFERASE GENE AND PROMOTER

(57) Abstract: The present invention provides a recombinant expression cassette comprising an  $\alpha$  1-3 galactosyltransferase promoter operably linked to a polynucleotide for expression. The invention also provides a recombinant mutating cassette comprising a region of homology to an  $\alpha$  1-3 galactosyltransferase genomic sequence. The cassettes can be employed to express foreign genes or to disrupt the native  $\alpha$  1-3 galactosyltransferase genomic sequence, particularly within an animal. Thus, the invention also provides transgenic animals and methods for their production and use.

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## $\alpha$ 1-3 GALACTOSYLTRANSFERASE GENE AND PROMOTER

### TECHNICAL FIELD OF THE INVENTION

This invention relates to the  $\alpha$ 1-3 galactosyltransferase gene, promoters  
5 therefor, and the use thereof to create transgenic animals.

### BACKGROUND OF THE INVENTION

The current shortage of acceptable organs for transplantation is a major health concern. Because the demand for acceptable organs exceeds the supply,  
10 many people die each year while waiting for organs to become available. To help meet this demand, research has been focused on developing alternatives to allogenic transplantation. Thus, for example, dialysis has been available to patients suffering from kidney failure, artificial heart models have been tested, and other mechanical systems have been developed to assist or replace failing organs.  
15 Such approaches, however, are quite expensive, and the need for frequent and periodic access to such machines greatly limits the freedom and quality of life of patients undergoing such therapy.

Xenograft transplantation represents a potentially attractive alternative to artificial organs for human transplantation. The potential pool of nonhuman  
20 organs is virtually limitless, and a successful xenograft transplantation would not render the patient virtually tethered to machines as is the case with artificial organ technology. Host rejection of such cross-species tissue, however, remains a major concern in this area. Some noted xenotransplants of organs from apes or old-world monkeys (e.g., baboons) into humans have been tolerated for months  
25 without rejection. However, such attempts have ultimately failed due to a number of immunological factors. Even with heavy immunosuppression to suppress hyperacute rejection, a low-grade innate immune response, attributable in part to failure of complement regulatory proteins (CRPs) within the graft tissue to control activation of heterologous complement on graft endothelium, ultimately leads to  
30 destruction of the transplanted organs (see e.g., Starzl, *Immunol. Rev.*, 141, 213-44 (1994)). In an effort to develop a pool of acceptable organs for xenotransplantation into humans, researchers have engineered animals producing human CRPs, an approach which has been demonstrated to delay, but not eliminate, xenograft destruction in primates (McCurry et al., *Nat. Med.*, 1, 423-27  
35 (1995); Bach et al., *Immunol. Today*, 17, 379-84 (1996)).

In addition to complement-mediated attack, human rejection of discordant xenografts appears to be mediated by a common antigen: the galactose- $\alpha$ (1,3)-

galactose (gal- $\alpha$ -gal) terminal residue of many glycoproteins and glycolipids (Galili et al., *Proc. Nat. Acad. Sci. (USA)*, 84, 1369-73 (1987); Cooper et al., *Immunol. Rev.*, 141, 31-58 (1994); Galili et al., *Springer Sem. Immunopathol.*, 15, 155-171 (1993); Sandrin et al., *Transplant Rev.*, 8, 134 (1994)). This antigen is  
5 chemically related to the human A, B, and O blood antigens, and it is present on many parasites and infectious agents, such as bacteria and viruses. Most mammalian tissue also contains this antigen, with the notable exception of old world monkeys and apes (including humans) (see Joziassse et al., *J. Biol. Chem.*, 264, 14290-97 (1989) and references cited therein)). The antigen is highly  
10 immunogenic in humans, and many individuals show significant levels of circulating IgG with specificity for gal- $\alpha$ -gal carbohydrate determinants (see, e.g., Galili et al., *J. Exp. Med.*, 162, 573-82 (1985), Galili et al., *Proc. Nat. Acad. Sci. (USA)*, 84, 1369-73 (1987)). Thus, in hopes of better understanding barriers to xenotransplantation, recent attention has turned to the enzyme mediating the  
15 formation of gal- $\alpha$ -gal moieties:  $\alpha$ 1-3 galactosyltransferase.

The expression of  $\alpha$ 1-3 galactosyltransferase is regulated both developmentally and in a tissue-specific manner. The cDNA for this enzyme has been isolated from many species, including pigs (Hoopes et al., poster presentation at the 1997 Xenotransplantation Conference, Nantes France; Katayama et al., *J. Glycoconj.*, 15(6), 583-99 (1998); Sandrin et al., *Xenotransplantation*, 1, 81-88  
20 (1994), Strahan et al., *Immunogenics*, 41, 101-05 (1995)), mice (Joziassse et al., *J. Biol. Chem.*, 267, 5534-41 (1992)), and cows (Joziassse et al., *J. Biol. Chem.*, 264, 14290-97 (1989)). While authors have proposed to eliminate the gene from xenograft donor animals (Sandrin et al. (1994), *supra*; U.S. Patent 5,821,117  
25 (Sandrin et al.)), gene knock-out procedures generally require knowledge of the genomic structure and sequence beyond the cDNA of a given gene. The genomic organization of the mouse  $\alpha$ 1-3 galactosyltransferase homologue has been deduced (Joziassse et al., *J. Biol. Chem.*, 267, 5534-41 (1992)), and human homologues are known to be inactive pseudogenes (see Joziassse et al., *J. Biol. Chem.*, 266, 6991-98 (1991); Larsen et al., *J. Biol. Chem.*, 265, 7055-61 (1990)).  
30 However, the genomic organization of an  $\alpha$ 1-3 galactosyltransferase homologue from a species that could serve as a xenograft donor for human recipients has yet to be deduced, and no promoter for any  $\alpha$ 1-3 galactosyltransferase homologue gene is known. As such, there exists a need for methods and reagents for  
35 facilitating xenotransplantation between species, particularly between species exhibiting differential expression of the gal- $\alpha$ -gal epitope.

### BRIEF SUMMARY OF THE INVENTION

The present invention provides a recombinant expression cassette comprising an  $\alpha$ 1-3 galactosyltransferase promoter operably linked to a polynucleotide for expression. The invention also provides a recombinant  
5 mutating cassette comprising a region of homology to an  $\alpha$ 1-3 galactosyltransferase genomic sequence. The cassettes can be employed to express foreign genes or to disrupt the native  $\alpha$ 1-3 galactosyltransferase genomic sequence, particularly within an animal. Thus, the invention also provides transgenic animals and methods for their production and use. These aspects of the  
10 invention, as well as additional inventive features, will be apparent from the accompanying drawing, sequence listing, and the following detailed description.

### BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A through 1I depict the genomic organization porcine  $\alpha$ 1-3  
15 galactosyltransferase gene. Figure 1A depicts all introns and exons of the gene, indicating the size of the respective elements. Figures 1B through 1I depict alternatively spiced variants isolated from pig aortic endothelial cells.

Figure 2 depicts the organization of a portion of the porcine  $\alpha$ 1-3 galactosyltransferase promoter.

20 Figure 3 depicts the organization of the alternate splicing patterns observed in the expression of the human untranslated  $\alpha$ 1,3 galactosyltransferase pseudogene.

### DETAILED DESCRIPTION OF THE INVENTION

25 In a first aspect, the present invention provides a recombinant expression cassette in which an  $\alpha$ 1-3 galactosyltransferase promoter is operably linked to a polynucleotide for expression. The expression cassette is "recombinant" in that within the inventive cassette, the polynucleotide for expression is other than one encoding  $\alpha$ 1-3 galactosyltransferase. The promoter and the polynucleotide are  
30 "operably linked" in that an event at the promoter (e.g., binding of cellular transcription factors and other DNA binding proteins) precipitates expression (i.e., transcription) of the polynucleotide. So long as this operable linkage is maintained, the cassette can include elements other than the  $\alpha$ 1-3 galactosyltransferase promoter and the polynucleotide for expression. For  
35 example, the cassette can contain polyadenylation sequences, repressors, enhancers, splice signals, signals for secretion (see, e.g., U.S. Patent 4,845,046 and European Patent EP-B-319,641), etc. Moreover, the expression cassette can



include more than one polynucleotide operably linked to the  $\alpha$ 1-3 galactosyltransferase promoter, (e.g., multiple coding sequences separated by internal ribosome entry sites).

The  $\alpha$ 1-3 galactosyltransferase promoter can be derived from any species normally expressing the gene. Thus, for example, the promoter can be derived from the bovine, porcine, or murine  $\alpha$ 1-3 galactosyltransferase genes. Examples of such promoters are set forth at SEQ ID Nos:1-6. However, the  $\alpha$ 1-3 galactosyltransferase promoter is not limited to one of these sequences, as it can be an active fragment of one of these sequences or a derivative of one of these sequences having one or more mutations (e.g., point mutations, substitutions, insertions, deletions, etc.). Furthermore, given the instant disclosure, it is within the ordinary skill of the art to assay regions of the  $\alpha$ 1-3 galactosyltransferase gene unrelated to SEQ ID NOs:1-6 for promoter activity, and the inventive expression cassette can include any  $\alpha$ 1-3 galactosyltransferase promoters so identified.

Suitable promoters can be readily identified by construction an expression cassette in which the derivative sequence is operably linked to a desired reporter gene (e.g., RNA for detection by Northern hybridization, or DNA encoding CAT, luciferase, green-fluorescent peptide,  $\beta$ -galactosidase, etc.) and introducing the cassette into a suitable environment for transcription and (where appropriate) translation.

Subsequently, promoter activity is detected by assaying for the presence of the reporter by standards methods (e.g., Northern hybridization, Southern hybridization, enzymatic detection, immunohistochemistry, etc.).

Within the expression cassette, the  $\alpha$ 1-3 galactosyltransferase promoter can be operably linked to any desired coding polynucleotide. Generally, where expression of a given gene or factor is desired, the skilled artisan will be in possession of the sequence of the coding polynucleotide. Thus, the polynucleotide can be expressed as a bioactive RNA molecule (e.g., an antisense RNA or a ribozyme). Alternatively, the polynucleotide can encode a protein of interest, and in this embodiment, the polynucleotide can be or comprise cDNA or genomic DNA.

Where the polynucleotide encodes a protein, any desired protein can be so encoded, and it need not be syngenic to the species from which the promoter is derived. Thus, for example, the cassette can be employed in animals to produce proteins facilitating growth or bulking of the animal (e.g., bovine or human growth factor) for conferring resistance to disease or parasites. Other encoded proteins can be enzymes such as sulfo- or glycosyltransferases, (e.g., a fucosyltransferase, a galactosidase, a galactosyltransferase, a  $\beta$ -acetylgalactosaminyltransferase, an

N-acetylglycosaminyltransferase, an N-acetylglucosaminyltransferase, a sialyltransferase, etc.). Where the expression cassette is employed to generate tissue or organs for xenotransplantation into an organism lacking gal- $\alpha$ -gal antigens (as described below), preferably the polynucleotide encodes a Type I  
5 fucosyltransferase, a Type II fucosyltransferase, an  $\alpha$  2-3 sialyltransferase, or an  $\alpha$  2-6 sialyltransferase from any species, the coding sequences of which are known (see, e.g., Larsen et al., *Proc. Nat. Acad. Sci. (USA)*, 87, 6674-78 (1990); Kelly et al., *J. Biol. Chem.*, 270(9), 4640-49 (1995), *J. Biol. Chem.*, 268(30), 22782-87 (1993), Weinstein et al., *J. Biol. Chem.*, 262(36), 17735-43 (1987)).

10 The expression cassette can be constructed by conventional methods of molecular biology (e.g., direct cloning by ligation, site specific recombination using recombinases, such as the flp recombinase or the cre-lox recombinase system (reviewed in Kilby et al. *Trends Genet.*, 9, 413-21 (1993)), homologous recombination, and other suitable methods). Typically, the promoter sequence is  
15 introduced into a vector 5' (i.e., "upstream") of the coding polynucleotide and any other elements (e.g., ribosome entry sites, polyadenylation sequences, etc.), after which the construct is subcloned and grown in a suitable host organism (e.g., yeast, bacteria, etc.) from which it can be isolated or substantially (and typically completely) purified by standard methods. Thus, the invention provides a vector  
20 (preferably an isolated or substantially purified vector) including a recombinant expression cassette as set forth above. Such a vector can be any desired type of vector, such as naked DNA vectors (e.g., oligonucleotides or plasmids); viral vectors (e.g., adeno-associated viral vectors (Berns et al., *Ann. N.Y. Acad. Sci.*, 772, 95-104 (1995)), adenoviral vectors (Bain et al., *Gene Therapy*, 1, S68  
25 (1994)), bacteriophages, baculovirus vectors (see, e.g., Luckow et al., *Bio/Technology*, 6, 47 (1988)), herpesvirus vectors (Fink et al., *Ann. Rev. Neurosci.*, 19, 265-87 (1996)), packaged amplicons (Federoff et al., *Proc. Nat. Acad. Sci. USA*, 89, 1636-40 (1992)), papilloma virus vectors, picornavirus vectors, polyoma virus vectors, retroviral vectors, SV40 viral vectors, vaccinia  
30 virus vectors) or other vectors (e.g., a cosmid, a yeast artificial chromosome (YAC), etc.). Of course, the vector can (and typically does) contain elements in addition to the expression cassette that are appropriate to the type of vector (e.g., origins of replication, marker genes, genes conferring resistance to antibiotics, etc.). The insertion of the expression cassette can disrupt one or more of these  
35 elements, if desired, or the cassette can be inserted between genetic elements to minimize perturbation of the backbone vector.

Where the vector is a viral vector, preferably it is replication incompetent. Thus, for example, an adenoviral vector preferably has an inactivating mutation in at least the E1A region, and more preferably in region E1 (i.e., E1A and/or E1B) in combination with inactivating mutations in region E2 (i.e., E2A, E2B, or both  
5 E2A and E2B), and/or E4 (see, e.g., International Patent Application WO 95/34671). An AAV vector can be deficient in AAV genes encoding proteins associated with DNA or RNA synthesis or processing or steps of viral replication (e.g., capsid formation) (see U.S. Patents 4,797,368, 5,354,768, 5,474,935, 5,436,146, and 5,681,731). Where the vector is a retroviral vector, the cis-acting  
10 encapsidation sequence (E) essential for virus production in helper cells can be deleted upon reverse transcription in the host cell to prevent subsequent spread of the virus (see, e.g., U.S. Patent 5,714,353). Where the vector is a herpesvirus, inactivation of the ICP4 locus and/or the ICP27 cassette renders the virus replication incompetent in any cell not complementing the proteins (see, e.g., U.S.  
15 Patent 5,658,724, see also DeLuca et al., *J. Virol.*, 56, 558-70 (1985); Samaniego et al., *J. Virol.*, 69(9), 5705-15 (1996)).

To use the inventive recombinant expression cassette, it is introduced into a eukaryotic cell in a manner suitable for the cell to express the coding polynucleotide. A vector harboring the recombinant expression cassette is  
20 introduced into a eukaryotic cell by any method appropriate for the vector employed, which generally are well-known in the art. Thus, plasmids are transferred by methods such as calcium phosphate precipitation, electroporation, liposome-mediated transfection, microinjection, viral capsid-mediated transfer, polybrene-mediated transfer, protoplast fusion, etc. Viral vectors are best  
25 transferred into the cells by infecting them.

Depending on the type of vector, it can exist within the cell as a stable extrachromosomal element (which can even be heritable, see e.g., Gassmann, M. et al., *Proc. Natl. Acad. Sci. (USA)*, 92, 1292 (1995)) or it can integrate into the host cell's chromosomes. Thus, the invention provides a chromosome including a  
30 recombinant expression cassette such as described above, as well as a cell including such a cassette (and such a chromosome). The  $\alpha$ 1-3 galactosyltransferase promoter of the expression cassette can be native to such a cell or chromosome, or it can be exogenous to the cell or chromosome. Where the promoter is native to the cell or chromosome, preferably the polynucleotide for  
35 expression within the cassette (the non-native polynucleotide) displaces the operable linkage between the native polynucleotide encoding  $\alpha$ 1-3 galactosyltransferase such that it is no longer operably linked to the native  $\alpha$ 1-3

galactosyltransferase promoter. Such displacement can be accomplished where the non-native polynucleotide is cloned between the promoter and the native polynucleotide (i.e., upstream of the native polynucleotide), especially where the non-native polynucleotide contains one or more transcriptional termination signals (preferably in all three putative reading frames). Of course, the non-native polynucleotide also can be introduced into the locus such that it destroys the native exon/intron boundaries and/or introduces inactivating mutations (e.g., deletions, insertions, frame-shifts, etc.) into the native coding sequence.

Preferably, the transgenic cell presents a suitable microenvironment for the coding polynucleotide within the expression cassette to be expressed. In many instances, the transgenic cells can be used to study the tissue specificity, dynamics, and kinetics of the promoter, for example by assaying for the expression of the polynucleotide within the cells. However, as the absence of activity is as useful as the presence of promoter activity in these contexts, any cell can be employed for such purposes; such a cell can be *in vivo* or *in vitro*. Preferably, the cell is derived from a species syngenic to the source of the promoter so that, by virtue of the properties of the  $\alpha$ 1-3 galactosyltransferase promoter present within the expression cassette, the polynucleotide within the cassette is expressed within such transgenic tissues, organs, or animals with the same kinetics and tissue specificity as the native  $\alpha$ 1-3 galactosyltransferase gene in wild-type animals. Where the cells are *in vivo*, they are typically cells of a mammal (e.g., human cells), and can be any type of cells. Suitable cells for use *in vitro* include yeast, protozoa (e.g., *T. cruzi* epimastigotes), cells derived from any mammalian species (e.g., VERO, CV-1, COS-1, COS-7, CHO-K1, 3T3, NIH/3T3, HeLa, C1271, BS-C-1 MRC-5, etc.), insect cells (e.g., *Drosophila* Snyder cells), or other such cells. In other applications, the cell can be employed to construct transgenic tissues, organs, or animals, as described below, in which case the cell typically is a spermatozoon, ovum, zygote, primordial germ cells, or embryonic stem cell.

In another embodiment, the invention provides a method of mutating a region of a chromosome comprising an  $\alpha$ 1-3 galactosyltransferase gene. In accordance with the inventive method, a recombinant mutating cassette comprising a region of homology to the  $\alpha$ 1-3 galactosyltransferase gene is recombined with a chromosome which has an  $\alpha$ 1-3 galactosyltransferase gene such that homologous recombination occurs between the cassette and the chromosome. As a result of the homologous recombination, a mutation is introduced into the native  $\alpha$ 1-3 galactosyltransferase chromosomal gene sequence.

Thus, the final step of the method involves screening for successful recombination.

The inventive method employs a recombinant mutating cassette including at least a first region of homology to an  $\alpha$ 1-3 galactosyltransferase genomic sequence, and the invention provides such a cassette. Within such a cassette, this  
5 first region of homology is adjacent to either to at least one polynucleotide for insertion or to a second region of homology. The mutating cassette is "recombinant" in that neither the second region of homology nor the polynucleotide for insertion is adjacent to the first  $\alpha$ 1-3 galactosyltransferase  
10 genomic sequence in its native state (i.e., within a chromosome).

The insertion cassette can include more than one polynucleotide for insertion and/or more than one region of homology to all or a portion of the  $\alpha$ 1-3 galactosyltransferase genomic sequence. Indeed, where the cassette includes a region for insertion, preferably it has at least two regions of homology flanking the  
15 region for insertion. Where more than one region of homology is present, whether adjacent to each other or flanking a region for insertion, the cassette can be used to replace any span of the target chromosomal genomic sequence that lies between the two homologous chromosomal regions. Where multiple regions of homology are present, they should generally be arrayed in the same 5' to 3' orientation  
20 relative to one another.

A region of homology can be homologous to any portion of the genomic sequence of an  $\alpha$ 1-3 galactosyltransferase gene or the antisense strand thereof. The region can be homologous to the gene of any desired species, such as those discussed above, and it can be homologous to an intron, an exon, a promoter  
25 sequence, or any other desired sequence from the genomic DNA. To this end, regions of homology can be selected from the promoter sequences disclosed in SEQ ID NOs:1-6. Alternatively (or additionally) a region of homology can be selected from a portion of the genomic sequence from an  $\alpha$ 1-3 galactosyltransferase homologue. In this light, some of the murine sequences have  
30 been published (see, e.g., Joziassse et al., *J. Biol. Chem.*, 267, 5534-41 (1992)), and additional portions are set forth as SEQ ID NOs: 17-25. Portions of the porcine genomic sequence are disclosed herein as SEQ ID NOs: 7-16. Portions of the human  $\alpha$ 1,3 galactosyltransferase pseudogene genomic sequences are set forth at SEQ ID NOs: 35-42, and various (untranslated) human cDNA transcripts are set  
35 forth as SEQ ID NOs: 27-34, and those from Rhesus monkeys are set forth at SEQ ID NOs: 43-44. These sequences disclosed herein, as well as the published murine sequences, include the intron/exon boundaries from which one of skill in

the art can isolate additional intronic genomic sequences by techniques such as genome walking, 5' RACE, 3' RACE, etc.

A region of homology to the genomic sequence of an  $\alpha$ 1-3 galactosyltransferase gene need not be an exact complement to the genomic sequence; however, the region must be sufficiently homologous to the  $\alpha$ 1-3 galactosyltransferase gene to permit homologous recombination between the cassette and the genomic DNA *in vivo*. Indeed, in some embodiments (e.g., for introducing point mutations into the genomic sequence), a region of homology preferably contains some mismatched bases. Thus, typically, the region of homology will bear at least about 75 % homology to a portion of the  $\alpha$ 1-3 galactosyltransferase gene or its antisense strand (such as at least about 85 % homology to a portion of the  $\alpha$ 1-3 galactosyltransferase gene or its antisense strand), and more typically the region of homology will bear at least about 90 % homology to a portion of the  $\alpha$ 1-3 galactosyltransferase gene or its antisense strand (such as at least about 95 % or even at least about 97 % homology to a portion of the  $\alpha$ 1-3 galactosyltransferase gene or its antisense strand). Any commonly employed method (e.g., BLAST database searching) for calculating percent homology can be used to select a suitable region of homology. Similarly, while the length of the region of homology is not critical, it should be sufficiently long to facilitate homologous recombination between the cassette and the genomic DNA *in vivo*. Thus, typically the region of homology will be at least about 50 nucleotides long (such as at least about 75 or 100 bases long), and more typically it will be at least several hundred bases long (such as at least about 250, 500, or even 750 bases long). Indeed, in many applications, the region of homology preferably is several thousand bases long to maximize the likelihood of homologous recombination *in vivo*. The ideal length of a region of homology depends in part on the number of such regions within the cassette – where one or few regions of homology are present, they should be longer to facilitate recombination between the cassette and the genomic DNA; conversely, where the cassette contains several regions of homology, they can be shorter without reducing the likelihood of recombination events.

Where present within the cassette, a region for insertion can be or comprise any DNA which is desired to be introduced into the genomic sequence of an  $\alpha$ 1-3 galactosyltransferase gene. Thus, the region can comprise genetic regulatory elements (e.g., enhancers, promoters, repressors, etc., the sequences of which are known) or consensus binding sites for DNA-binding proteins (e.g., restriction endonucleases, transcription factors, etc.). In many applications, a region for

insertion can comprise a polynucleotide for expression, such as those set forth above, or even expression cassettes. A preferred polynucleotide for insertion is an expression cassette for expressing a positive marker flanked by FRT sites, thus facilitating the identification of chromosomes into which the polynucleotide for  
5 insertion has integrated as well as excision of the cassette.

The mutating cassette can be constructed by any desirable molecular techniques, and typically, the mutating cassette will be engineered within a vector, such as those set forth above. Typically, the vector is a gene transfer vector suitable for introducing the cassette into a host cell. In addition to the region(s) of  
10 homology and the polynucleotide for insertion elements, the mutating cassette can have other components, such as, for example, an expression cassette, a region of homology to other genes or chromosomal regions, a polyadenylation sequence, etc., and it is preferred that the insertion cassette comprises a cassette for expressing at least one marker gene (which may be or comprise the polynucleotide  
15 for insertion). Such a marker can be either positive (conferring a visible phenotype to the cells) or negative (killing cells or rendering non-recombinant cells growth-impaired), and both can be used in conjunction. Examples of such positive and negative selection markers are the neosporin resistance ( $neo^R$ ) gene, the hydromycin resistance ( $hyg^R$ ) gene, and a thymidine kinase gene (e.g., HSV  
20 tk); other suitable markers are known in the art (see, e.g., Mansour et al., *Nature*, 336, 348-52 (1988); McCarrick et al., *Transgen. Res.*, 2, 183-90 (1993)). A marker gene sequence can be bordered at both ends by FRT DNA elements, and/or with stop codons for each of the three putative reading frames being inserted 3' to the desired DNA sequence. Presence of the FRT elements permits the marker to  
25 be deleted from the targeted chromosome, and the stop codons ensure that the  $\alpha 1,3$  galactosyltransferase gene remains inactivated following deletion of the selectable marker, if inactivation is the desired result of the use of the mutating cassette. The relative orientations of the positive and negative selectable markers are not critical. However, where a positive marker is employed, it should be  
30 located between regions of homology, while any negative marker should be outside the regions of homology, either 5' or 3' to those regions.

In accordance with the inventive method, homologous recombination occurs between the  $\alpha 1,3$  galactosyltransferase genomic chromosomal DNA and the region (or regions) of homology in the mutating cassette. Where more than  
35 one region of homology is present in the cassette, any portion of the genome lying between the homologous target sequences is replaced by whatever sequence lies between the regions of homology in the cassette. Thus, where the mutating

cassette contains a region for insertion flanked by two regions of homology, it will be introduced into the genomic sequence adjacent to the sites of homology, replacing that portion of the genomic sequence. Of course, where the two flanking regions of homology are normally adjacent to each other in the chromosomal  
5 sequence, the region for insertion is introduced into the chromosome without replacing any native sequence. Similarly, where no region for insertion is present within the cassette, that portion of the chromosome lying between the two regions of homology in the cassette is deleted as a result of the recombination events. Where the cassette contains a region of homology that differs slightly from the  
10 homologous sequence within the genome, it can be employed to introduce point mutations into the genomic sequence.

While the recombination event can occur *in vitro*, typically such homologous recombination occurs within a host cell between an exogenous vector containing the cassette and a chromosome within the host cell containing an  $\alpha$ 1-3  
15 galactosyltransferase genomic sequence. Thus, the present invention provides a cell harboring a mutating cassette, as described above. The vector can be introduced into the host cell by any appropriate method, such as set forth above. Commonly, however, the vector is introduced into small cells (e.g., embryonic stem cells) by electroporation and into large cells (e.g., ova or zygotes) by  
20 microinjection. Where microinjection is employed, the vector preferably is injected directly into a nucleus or pronucleus of the cell.

The last step in the method is to screen for successful recombination events. Any assay to detect such events can be employed in the context of the inventive method. In accordance with one such assay, chromosomal DNA is  
25 screened by PCR or Southern hybridization. For example, where the mutating cassette is designed to delete a portion of the  $\alpha$ 1-3 galactosyltransferase genomic sequence, the absence of signal using a probe or primer directed against the region to be deleted indicates a positive recombination event. Conversely, where the cassette includes a region for insertion, a positive result using a probe or primer  
30 directed against the region for insertion is indicative of a positive recombination event. Of course, the chromosomal DNA can be sequenced to confirm the correct insertion/deletion/replacement. Where recombination is directed within cells, the events can be screened by assaying for any markers present in the mutating cassette.

35 By employing the inventive method, one of skill in the art can use the inventive mutating cassette to introduce targeted deletions, insertions, or replacement mutations into any predefined site within the  $\alpha$ 1-3



galactosyltransferase genomic sequence. Any desired amount or portion of the gene can be thus deleted, which can lead to complete inactivation of the gene. For introducing inactivating mutations into the gene, preferably at least one region of homology is selected to recombine with the promoter (to inactivate it) or exons 4-9, which contain the coding sequences. Similarly, the inventive method can introduce functional expression cassettes in place of the  $\alpha$ 1-3 galactosyltransferase gene, which can be under the control of the native  $\alpha$ 1-3 galactosyltransferase promoter or an exogenous promoter within the cassette (especially where the native  $\alpha$ 1-3 galactosyltransferase promoter is destroyed). Thus, the present invention provides a recombinant chromosome containing such a mutation, and a recombinant cell comprising such a chromosome.

As mentioned above, the invention provides recombinant cells and chromosomes comprising a recombinant expression cassette comprising an  $\alpha$ 1-3 galactosyltransferase promoter or a mutating cassette, as described above. Indeed, as a result of using these reagents and methods, the invention also provides a cell having a mutant  $\alpha$ 1-3 galactosyltransferase genomic sequence, as described above. While any cell having such exogenous genetic sequences is within the scope of the invention, preferably the cells are suitable for constructing a recombinant animal, and are most preferably totipotent cells. Thus, preferred cells are embryonic stem (ES) cells, ova, primordial germ cells (PGCs), and zygotes. ES cells and PGCs are especially preferred because such cells can be obtained and cultured in relatively large numbers relative to ova and zygotes. Using such cells, a transgenic animal having an expression cassette comprising an  $\alpha$ 1-3 galactosyltransferase promoter or a disruption in this gene can be constructed by methods known in the art (see e.g., U.S. Patents 5,850,004 (MacMicking et al.), 5,942,435 (Wheeler), 5,523,226 (Wheeler), and 5,175,383; White et al., *Transplant. Int.*, 5, 648-50 (1992); McCurry et al., *Nat. Med.*, 1, 423-427 (1995); Hogan et al., *Manipulating the Mouse Embryo*, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1986); Hammer et al., *Nature*, 315, 680 (1985); Murray et al., *Reprod. Fert. Devl.*, 1, 147, (1989); Purselet et al., *Vet. Immunol. Histopath.*, 17, 303 (1987); Rexroad et al., *J. Reprod. Fert.*, 41, (suppl.), 119 (1990); Rexroad et al., *Molec. Reprod. Devl.*, 1, 164 (1989); Simonset al., *BioTechnology*, 6, 179 (1988); Vize et al., *J. Cell. Sci.*, 90, 295 (1988); Wagner, J. *Cell. Biochem.*, 13B (suppl.), 164 (1989); Thomas et al., *Cell*, 51, 503 (1987); Capecchi, *Science*, 244, 1288 (1989); Joyner et al., *Nature*, 338, 153 (1989); Ausubelet al., *Cur. Prot. Mol. Biol.*, John Wiley & Sons (1987)).

Where ova and zygotes are employed, after the introduction of the cassette, they can be implanted into surrogate mothers to develop into adult animals. Where ES cells or PGCs are employed, after the introduction of the cassette, they typically are further manipulated (e.g., by injection into a blastocyst or morula, co-culture with a zona pellucida-disrupted morula, fusion with an enucleated zygote, etc.) such that their mitotic descendants are found in a developing embryo. Such an embryo typically is a chimera composed of normal embryonic cells as well as mitotic descendants of the introduced ES cells or PGCs. Alternatively, the genome of an ES cell or PGC can be incorporated into an embryo by fusing the ES cell/PGC with an enucleated zygote to create a non-chimeric embryo in which all nuclei are mitotic descendants of the fused ES cell/PGC nucleus. In any event, to produce a transgenic animal, the embryo or zygote is implanted into a pseudopregnant animal, which, after suitable gestation, gives birth to an animal containing the mutant chromosome containing the cassette in its germ line (if a chimera) or possibly all of its cells. Of course, as mentioned above, where the animal is engineered to include a non-mutating expression cassette, it can be inherited as an extrachromosomal plasmid (Gassmann, M. et al., *supra*). However constructed, the presence of the recombinant allele can be confirmed by performing Northern hybridization or rt-PCR on RNA isolated from the animal in question.

After birth and sexual maturation, a chimeric animal can be mated to generate a heterozygous animal comprising a disrupted  $\alpha$ 1-3 galactosyltransferase gene or recombinant expression cassette (integrated or extrachromosomal) including a  $\alpha$ 1-3 galactosyltransferase promoter. Heterozygotes can be crossed to produce a homozygous strain. Such animals having a recombinant expression cassette including an  $\alpha$ 1-3 galactosyltransferase promoter, as discussed above, will express the polynucleotide for expression of such cassette within the same tissue types and with the same kinetics as a wild-type animal of the same species and strain expresses the  $\alpha$ 1-3 galactosyltransferase gene. Of course, homozygous transgenic animals of the present invention having a disruption in the  $\alpha$ 1-3 galactosyltransferase gene will produce altered forms of the protein or no functional protein at all. Desirably, the phenotype of such "knock out" animals relative to an animal having a wild type  $\alpha$ 1-3 galactosyltransferase gene is a markedly increased time of survival of cells isolated or derived from the transgenic animal in the presence of human serum, which can be assessed by any desired method (see, e.g., Osman et al., *Proc. Nat. Acad. Sci. (USA)*, 94, 14677-82 (1997)).

The inventive transgenic animals are useful for any use to which animals can be put, and they can be any desired species (e.g., pigs, cows, mice, cats, dogs, etc.). Transgenic mice in which a reporter gene is operably linked to the  $\alpha$ 1-3 galactosyltransferase promoter are valuable reagents for assessing the activity and specificity of the promoter. Transgenic livestock (e.g., pigs, cows, goats, and the like) having an inventive expression cassette in which a growth hormone is expressed under the control of the  $\alpha$ 1-3 galactosyltransferase promoter can be matured or bulked better than commonly employed strains. Tissue obtained from a transgenic animal according to the present invention can be implanted into a host according to standard surgical methods, and the invention concerns a method of xenotransplantation from a transgenic animal as described herein. The invention also provides a transgenic organ consisting essentially of transgenic cells engineered as described above (e.g., a lung, a heart, a liver, a pancreas, a stomach, an intestine, a kidney, a cornea, skin, etc.), particularly for use in the method of transplantation. The host can be any animal host, such as a pig, a dog, a cat, a cow, a goat, etc. Of course, the recipient can be a human as well, in which case the source animal preferably is a pig.

Transgenic animals lacking a functional  $\alpha$ 1-3 galactosyltransferase gene are attractive sources of organs and tissues for xenotransplantation into primates, especially humans, because the tissues of such animals lack the highly antigenic gal- $\alpha$ -gal epitope. Similarly, transgenic pigs having a recombinant expression cassette in which a coding sequence for Type I fucosyltransferase, a Type II fucosyltransferase (especially  $\alpha$ (1,2) fucosyltransferase), an  $\alpha$  2-3 sialyltransferase, or an  $\alpha$  2-6 sialyltransferase is operably linked to the  $\alpha$ 1-3 galactosyltransferase promoter also are suitable sources of xenotransplantation tissues, as these encoded enzymes compete for the same substrate as  $\alpha$ 1-3 galactosyltransferase, and their presence can reduce (preferably below an antigenic threshold) the gal- $\alpha$ -gal antigens in tissues derived from such animals. Indeed,  $\alpha$ (1,2) fucosyltransferase converts this substrate into the universally-tolerated H antigen (i.e., the "O" blood-type antigen) and also blocks the addition of the  $\alpha$ 1,3 gal moiety. As such, a gene encoding  $\alpha$ (1,2) fucosyltransferase is an especially preferred polynucleotide for expression to be included within the inventive recombinant expression cassette. A preferred source animal for xenotransplantation tissues (and by extension the tissues themselves) preferably contains a disruption in the  $\alpha$ 1-3 galactosyltransferase gene as well as having a recombinant expression cassette in which a coding sequence for Type I fucosyltransferase, a Type II fucosyltransferase (especially  $\alpha$ (1,2)

fucosyltransferase), an  $\alpha$  2-3 sialyltransferase, or an  $\alpha$  2-6 sialyltransferase is operably linked to the  $\alpha$ 1-3 galactosyltransferase promoter. More preferably, the animal contains a disruption in the native promoter of  $\alpha$ 1-3 galactosyltransferase and an  $\alpha$ (1,2) fucosyltransferase coding sequence under the control of its own  
5 promoter. Most preferably, the source animal also expresses exogenous human complement regulatory proteins, as discussed above, to further minimize host resistance of the xenograft tissue.

It will be apparent that a transgenic animal created in accordance with the invention can have the exogenous gene cloned in place of the native  $\alpha$ 1,3  
10 galactosyltransferase gene (i.e., a “knock-in” approach). Indeed, in many embedment such a “knock-in” approach is preferable, for example to avoid the potential of the development of congenital cataracts in purely “knock-out” animals (e.g., as a result of opportunistic infections of microbes bearing the gal- $\alpha$ -gal motif). Indeed, such an approach can afford a safe alternative to broadband  
15 antibiotics in livestock and pets, a current public health concern. In this respect, the invention can be employed to create heartier and healthier livestock and pets.

While one of skill in the art is fully able to practice the instant invention upon reading the foregoing detailed descriptions, in conjunction with the drawing and the sequence listing, the following examples will help elucidate some of its  
20 features. In particular, these examples indicate how the genomic structure of the porcine  $\alpha$ 1-3 galactosyltransferase gene is elucidated, and how the identity and activity of the  $\alpha$ 1-3 Galactosyltransferase promoter is assessed. As these examples are presented for purely illustrative purposes, they should not be used to construe the scope of the invention in a limited manner, but rather should be seen  
25 as expanding upon the foregoing description of the invention as a whole.

Many experiments described in these examples employed well known techniques and reagents (see, e.g., Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2d edition, Cold Spring Harbor Press (1989)). Accordingly, in the interest of brevity, the examples do not present the experimental protocols in  
30 detail. In the experiments, enzymatic isolation and culture of porcine aortic endothelial cells (PAEC) was performed. PAEC were maintained in Dulbecco's modified essential medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 10,000 units of Heparin (ELKINS-SINN, Inc., Cherry Hill, NJ), 15 mg of endothelium growth supplement (Collaborative Biomedical Product Inc., Bedford,  
35 MA), L-glutamine, and penicillin-streptomycin. RNA was obtained from the organs of pigs (Brain, Heart, Spleen, Gut, and Thymus) and PAEC using Trizol reagent

(Gibco Ltd.,). Primers used to clone and identify regions of the porcine, murine, human, and Rhesus monkey genes are set forth at SEQ ID NOs: 45-96.

### Example 1

5 This example describes the identification of the 5' untranslated region and genomic structure of the porcine  $\alpha$ 1-3 galactosyltransferase gene.

A comparison of published sequences for the  $\alpha$ 1-3 galactosyltransferase cDNA (Hoopes et al., *supra*, Katayama et al., *supra*; Sandrin et al., *supra*; and Strahan et al., *supra*) revealed a divergence in the 5' boundary. Some of these  
10 cDNA contain putative 5' untranslated sequences that bear a high (> 70 %) homology to murine sequences identified as the second exon, and it was hypothesized that this region is conserved as an exon in the porcine genome as well.

Further 5' sequence was cloned using 5' RACE, and the putative  
15 transcription initiation site was probed by S1 protection assay, using standard protocols. Briefly, a plasmid containing the upstream genomic sequence was digested with restriction enzyme, Pml I, and linearized. The DNA was phosphorylated with shrimp alkaline phosphatase, heated to inactivate the enzyme, and then precipitated with ethanol. The linearized plasmid was digested again  
20 with Bgl II to yield a probe fragment, which was then end-labeled with  $\gamma$ -<sup>32</sup>P-ATP.

The probe was purified using G-25 sephadex, and about 16  $\mu$ l was mixed with 20  $\mu$ g of total RNA from pig aortic endothelial cells (PAEC), pig brain, and yeast (control), and the aliquots were coprecipitated using NH<sub>4</sub>OAc and ethanol. Pellets were resuspended in a standard hybridization buffer, heated to 95 °C for 3-  
25 4 minutes, and then incubated at 42 °C overnight.

After incubation, the yeast sample was split into two aliquots, and to each was added a standard S1 nuclease buffer. S1 nuclease was added to one aliquot, while the other did not receive the enzyme. The PAEC and brain samples each received the enzyme and the buffer. All samples were incubated for 30 minutes at  
30 37 °C, after which the reactions were stopped by the addition of a standard S1 inactivation buffer. Following the reaction, the samples were then precipitated, resuspended in 5  $\mu$ l of a standard gel loading buffer, and resolved using a 6% denaturing polyacrylamide gel.

The data revealed at least 8 separate alternatively spliced transcripts from  
35 PAEC, and additional splicing patterns from brain transcripts. Analysis of these sequences revealed three potential upstream exons (1, 1A, and 2), the boundaries of which comply with the AG-GT consensus, and six coding exons (4-9) also were

identified, which agreed with published results. Interestingly, the pig sequence seemingly lacks upstream exon 3 of the mouse 5' untranslated region. The overall organization of the pig genome is depicted in Figure 1. Alternatively spliced forms isolated from PAED are indicated in Figures 1B through 1I. Exon 1A is  
5 observed in transcripts isolated from brain tissue.

As mentioned, the transcripts obtained from PAEC and brain revealed several alternative splicing patterns. Using the genomic clone, intronic sequences were identified by "gene walking" using the method and reagents supplied with the UNIVERSAL GENEOMEWALKER™ KIT (Clontech Labs., Inc.). Primers  
10 (Seq ID NOs:41-56) were designed to hybridize with the cDNA, and also to the adapter sequence supplied with the Clontech kit. A series of nested PCR reactions was then performed to clone SEQ ID NOs:7-16, which were sequenced. From these results, the intron/exon boundaries were elucidated.

Summing the nucleotides of all identified exons predicts a transcript of  
15 about 3.8 kb. This prediction was assessed by Northern analysis. 20 µg of total RNA from PAEC, and pig brain, heart, spleen, gut, and thymus, were respectively separated on formamide agarose gels, and electrotransferred onto nylon membrane. The blots were hybridized with radiolabeled probes ( $2.5-4.0 \times 10^4$  cpm/ml) specific for pig GT exon 1 and exon 9 identified. The blots were exposed  
20 to Bio-MAX films (Eastman Kodak Co., Rochester, NY) for 6 days with intensifying screen. The results revealed primary transcripts of between 3.5-3.8 kb, in accordance with the predicted size and the published size for the bovine transcript.

## 25 **Example 2**

This example describes the identification of the 5' untranslated region and organization of the murine  $\alpha 1$ -3 galactosyltransferase gene.

To identify the 5' and 3' ends of  $\alpha 1,3$ GT gene transcripts, 5'- and 3'-RACE procedures were performed using the Marathon cDNA Amplification Kit  
30 (Clontech) with the spleen poly A<sup>+</sup> RNA of Balb/C adult male as template. To identify exon-intron boundaries or 5'- and 3'-flanking region of the transcripts, Murine GenomeWalker libraries were constructed using the Universal GenomeWalker Library Kit (Clontech) with Balb/C genomic DNA.

The results of these experiments revealed several genomic sequences,  
35 which are set forth at SEQ ID NOs: 17-25. The deduced 5' untranslated nucleotide sequences are longer by 56 bp than previously reported (Joziasse et al., *J. Biol. Chem.*, 267, 5534-41 (1992)). The relative intensity of Luciferase activity

by the pGL3/1280 construct was 15-fold higher than that of pGL3-Basic. The 3'-RACE revealed an extended 3'-UTR sequence 30 bp more than previously reported (*Id.*), but no other 3' UTR exon usage. The overall length of the transcript was 2586 bp, 89 bp longer than previously reported (*Id.*).

- 5           An overall comparison of 5'-UTR of cDNA sequences of the porcine (747 bp) and murine (492 bp)  $\alpha$ 1,3GT gene indicates that the homology is observed only in the region of exon 2 (71.7%). Exon 3 observed in mice is not observed in the pig. Murine exon 1 shows no homology with porcine exon 1.

### 10   **Example 3**

This example describes the identification of the organization of the human and Rhesus monkey  $\alpha$ 1-3 galactosyltransferase untranslated pseudogene.

- Working from published partial sequence of the human  $\alpha$  1,3 GT ninth exon, primers were designed to identify the start and end of the gene by 5'-RACE, 3'RACE and rPCR, as described above. Several alternate transcripts were identified, and these are set forth as SEQ ID NOs:27-34. The sequences were compared to those of other species employing a formula based on the consensus motif of the splicing acceptor junction: total number of pyrimidines plus 1 (for a branched A) among forty nucleotides per junction. Intron exon boundaries were confirmed as discussed above (see SEQ ID NOs: 35-42). The organization of the alternative splicing patterns observed is indicated in Figure 3.

- Using similar techniques, primers were designed based on a partial published sequence (Genbank Accession No. M73306) having homology to exon 9. Initially, 3'RACE showed only poly-A tails, evidence that transcripts exist. 5'-RACE results revealed sequences of high homology to those  $\alpha$ 1,3 sequences previously identified (e.g., porcine, bovine and murine), consistent with the identity of the sequence as the Rhesus pseudogene. The sequence of the Rhesus monkey transcripts are set forth at SEQ ID NOs: 43 and 44.

### 30   **Example 4**

This example describes the identification of the porcine, murine, and bovine  $\alpha$ 1-3 galactosyltransferase promoters.

- Using PCR and restriction digestions, various sized fragments between nucleotides 1981 and 2992 of SEQ ID NO:7 (porcine) and between nucleotides 375 and 1325 (murine) were generated. The fragments were cloned into a plasmid such that they were operably linked to a luciferase coding sequence. PAEC were then transfected with these constructs and probed for luciferase activity, along

with a positive and a negative (no promoter) control. All fragments exhibited significantly greater promoter activity over the negative control (between about 15% and 90 % relative light units, as compared to the positive control, the negative control exhibiting no luciferase activity). These results indicate that the regions are promoters and that the 5'-RACE results discussed in Examples 1 and 2 most likely represent the potential transcription initiation site (TIS). Moreover, sequence analysis of these regions reveals the presence of at least 8 SP1 or GC boxes within it and potentially seven AP-2 consensus binding motifs (see also Figure 2). This suggests that the gene may contain alternative start sites, and that sequences within exon 1 may also contain promoter activity. Other sequences from which  $\alpha$  1,3 GT promoters can be derived are set forth as SEQ ID NOs:1-6.

All of the references cited herein, including patents, patent applications, and publications, are hereby incorporated in their entireties by reference.

While this invention has been described with an emphasis upon preferred embodiments and illustrative examples, it will be obvious to those of ordinary skill in the art that variations of the preferred embodiments may be used and that it is intended that the invention may be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the invention as defined by the following claims.



## WHAT IS CLAIMED IS:

1. A recombinant expression cassette comprising an  $\alpha$ 1-3 galactosyltransferase promoter operably linked to a polynucleotide for expression, other than a polynucleotide encoding  $\alpha$ 1-3 galactosyltransferase.
- 5        2. The recombinant expression cassette of claim 1, wherein said promoter is derived from the bovine, porcine, or murine  $\alpha$ 1-3 galactosyltransferase genes.
3. The recombinant expression cassette of claim 1 or 2, wherein said promoter comprises any of SEQ ID Nos:1-6.
4. The recombinant expression cassette of any of claims 1-3, wherein said  
10 promoter comprises an active derivative of any of SEQ ID Nos:1-6.
5. The recombinant expression cassette of any of claims 1-4, wherein said polynucleotide for expression encodes an antisense RNA molecule or a ribozyme.
6. The recombinant expression cassette of any of claims 1-5, wherein said polynucleotide for expression encodes a protein.
- 15        7. The recombinant expression cassette of claim 6, wherein said protein is a fucosyltransferase, a galactosyltransferase, a  $\beta$ -acetylgalactosaminyltransferase, an N-acetylglycosaminyltransferase, an N-acetylglucosaminyltransferase, a sialyltransferase, or a sulfotransferase.
8. The recombinant expression cassette of claim 6, wherein said protein is a  
20 Type I fucosyltransferase, a Type II fucosyltransferase, an  $\alpha$  2-3 sialyltransferase, or an  $\alpha$  2-6 sialyltransferase.
9. The recombinant expression cassette of any of claims 6-8, wherein said polynucleotide for expression is heterogenic to said promoter.
10. The recombinant expression cassette of claim 9, wherein said  
25 polynucleotide for expression is human and wherein said promoter is porcine.
11. The recombinant expression cassette of any of claims 6-8, wherein said polynucleotide for expression is a cDNA.
12. The recombinant expression cassette of any of claims 6-8, wherein said polynucleotide for expression is genomic DNA.
- 30        13. A recombinant mutating cassette comprising a first region of homology to an  $\alpha$ 1-3 galactosyltransferase genomic sequence adjacent to either a second region of homology to said  $\alpha$ 1-3 galactosyltransferase genomic sequence or a polynucleotide for insertion.
14. The recombinant mutating cassette of claim 13, comprising first and  
35 second regions of homology to an  $\alpha$ 1-3 galactosyltransferase genomic sequence flanking a polynucleotide for insertion.

15. The recombinant mutating cassette of claim 13 or 14, wherein a region of homology is homologous to an exon, an intron, or a promoter of said  $\alpha$ 1-3 galactosyltransferase genomic sequence.

16. The recombinant mutating cassette of any of claims 13-15, wherein a  
5 region of homology is homologous to all or a portion of any one of SEQ ID NOs: 1-42.

17. The recombinant mutating cassette of any of claims 13-16, wherein said polynucleotide for insertion comprises an expression cassette.

18. The recombinant mutating cassette of claim 17, wherein said  
10 expression cassette encodes a marker.

19. A vector comprising the recombinant cassette of any of claims 1-18.

20. The vector of claim 19, which is an oligonucleotide, a plasmid, a cosmid, or a virus.

21. A transgenic cell harboring the vector of claim 19 or 20.

15 22. A chromosome comprising the recombinant expression cassette of any of claims 1-18.

23. A transgenic cell harboring the chromosome of claim 22.

24. The transgenic cell of claim 23, wherein said  $\alpha$ 1-3 galactosyltransferase promoter is native to said cell.

20 25. The transgenic cell of claim 23 or 24, wherein said polynucleotide for expression displaces a native polynucleotide encoding  $\alpha$ 1-3 galactosyltransferase.

26. The transgenic cell of claim 23 or 24, wherein said polynucleotide for expression is cloned between said promoter and a native polynucleotide encoding  $\alpha$ 1-3 galactosyltransferase.

25 27. The transgenic cell of claim 26, wherein said polynucleotide for expression comprises a stop codon.

28. The transgenic cell of any of claims 21, or 23-27, which is an embryonic stem cell, an ovum, a primordial germ cell, a spermatozoon, or a zygote.

30 29. The transgenic cell of any of claims 21, or 23-27, which expresses said polynucleotide for expression.

30. The cell of claim 29, wherein said polynucleotide for expression encodes a Type I fucosyltransferase, a Type II fucosyltransferase, an  $\alpha$  2-3 sialyltransferase, or an  $\alpha$  2-6 sialyltransferase, and wherein said cell produces said  
35 protein.

31. The transgenic cell of any of claims 21 or 23-30, wherein said cell produces a heterogenic complement regulatory protein (CRP).

32. The transgenic cell of claim 31, wherein said CRP is human and wherein said cell is nonhuman.

33. An embryo consisting essentially of transgenic cells according to any of claims 21 or 23-32.

5        34. An organ consisting essentially of transgenic cells according to any of claims 21 or 23-32.

35. The organ of claim 34, which is a lung, a heart, a liver, a pancreas, a stomach, an intestine, a kidney, or skin.

10       36. A transgenic animal consisting essentially of transgenic cells according to any of claims 21 or 23-32.

37. The transgenic animal of claim 30, which is a cattle, a mouse, a pig, a cat or a dog.

15       38. A transgenic knockout animal comprising a homozygous disruption in an endogenous  $\alpha$ 1-3 galactosyltransferase gene, wherein said disruption prevents the expression of a functional  $\alpha$ 1-3 galactosyltransferase protein.

39. The transgenic knockout animal of claim 38, wherein cells isolated from said knockout animal exhibit an increased time of survival in the presence of human serum relative to comparable cells isolated from an animal having a wild type  $\alpha$ 1-3 galactosyltransferase gene.

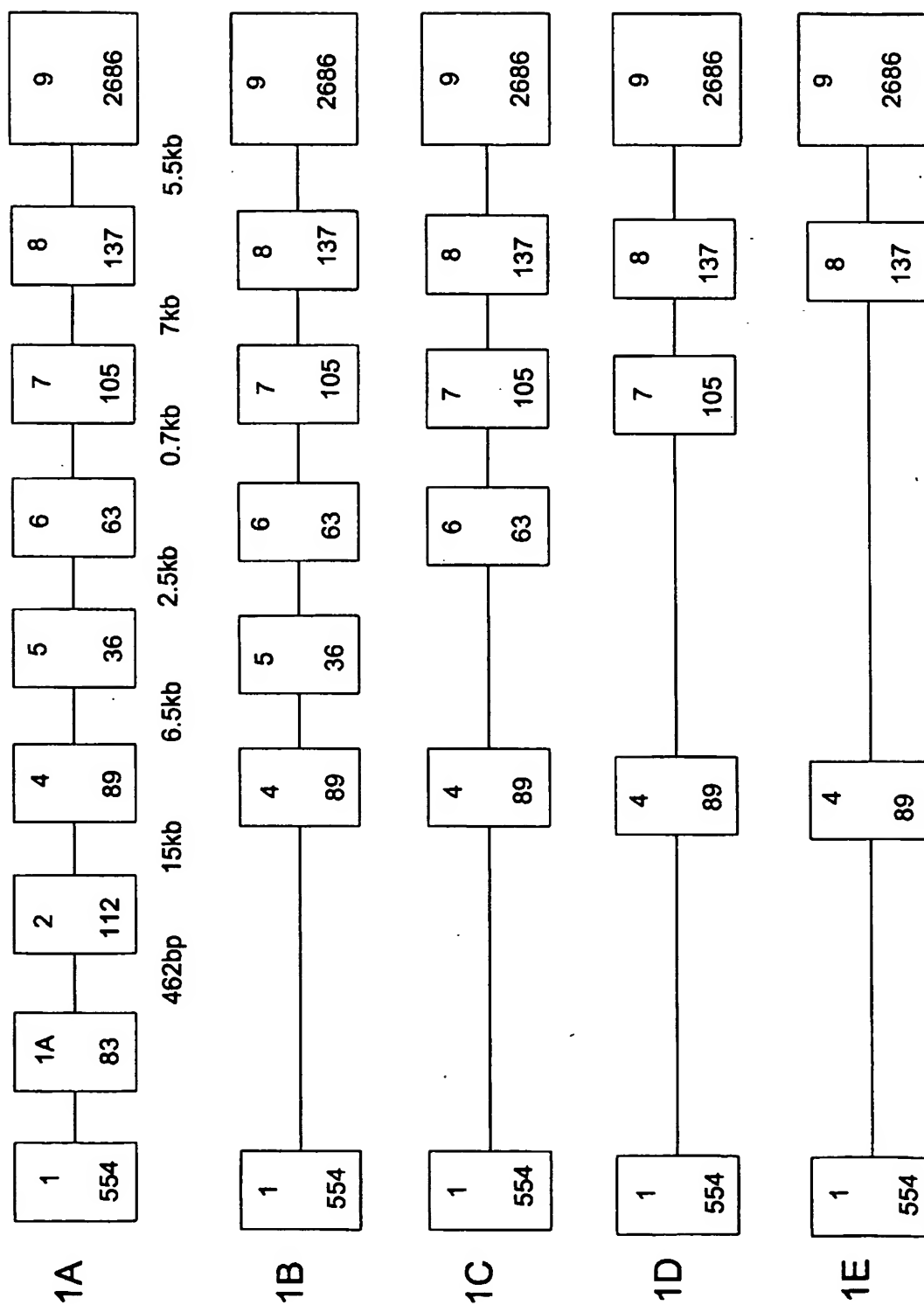
20       40. The transgenic knockout animal of claim 38 or 39, wherein the insertion replaces DNA at the start of the coding region of said  $\alpha$ 1-3 galactosyltransferase protein.

41. The transgenic knockout animal of claim 38 or 39, wherein the insertion replaces the promoter of said wild type  $\alpha$ 1-3 galactosyltransferase gene.

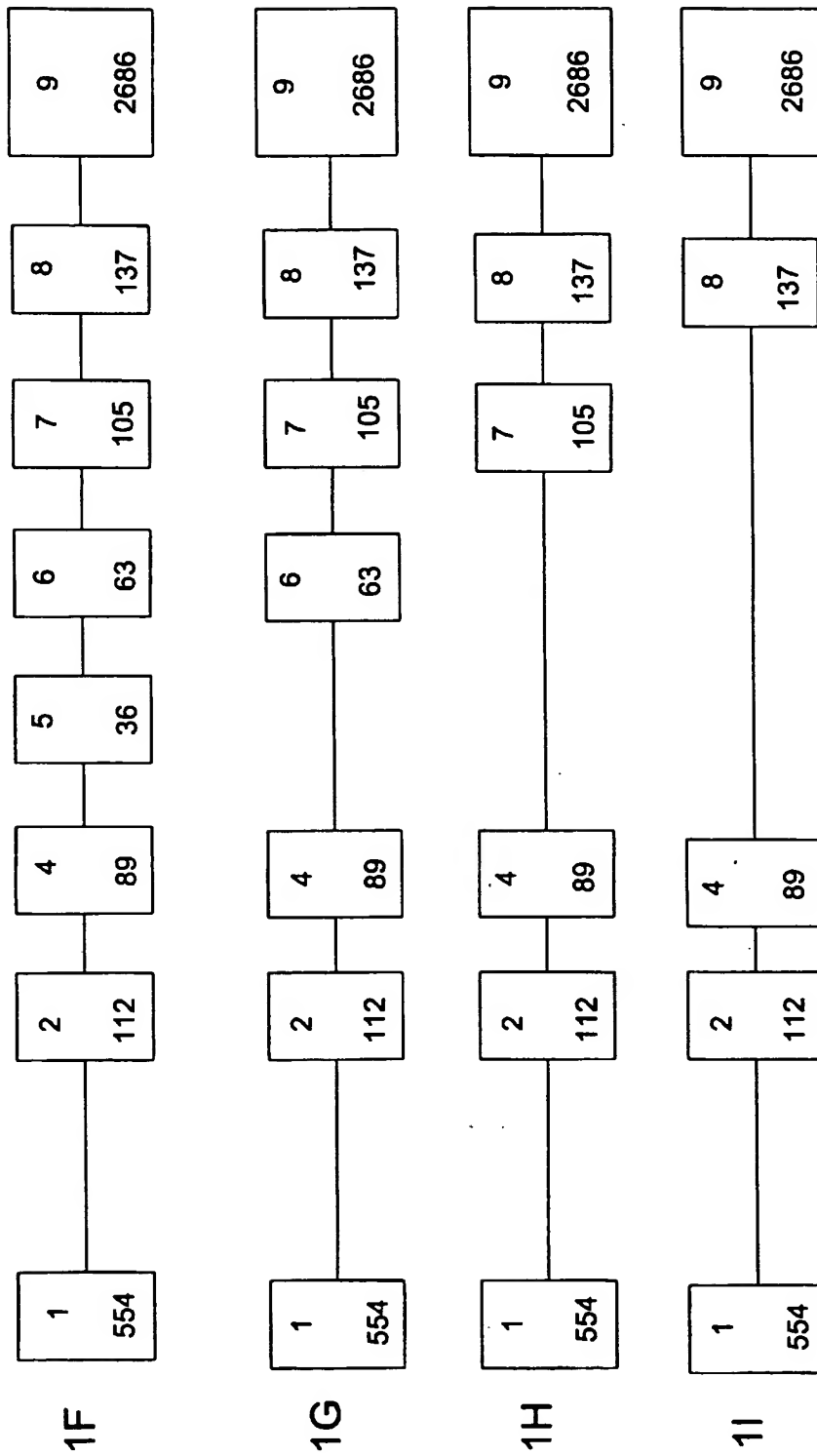
25       42. The transgenic knockout animal of any of claims 38-41, which produces at least one human protein selected from the group of proteins consisting of  $\alpha$ 1-3 galactosyltransferase,  $\alpha$ (1,2) fucosyltransferase, and complement regulatory proteins.

30       43. The transgenic knockout animal of any of claim 38-42, which is a pig.

1 / 4



2 / 4



E/G.2

4 / 4

Schematic genomic organization of human  $\alpha 1, 3$ GT gene

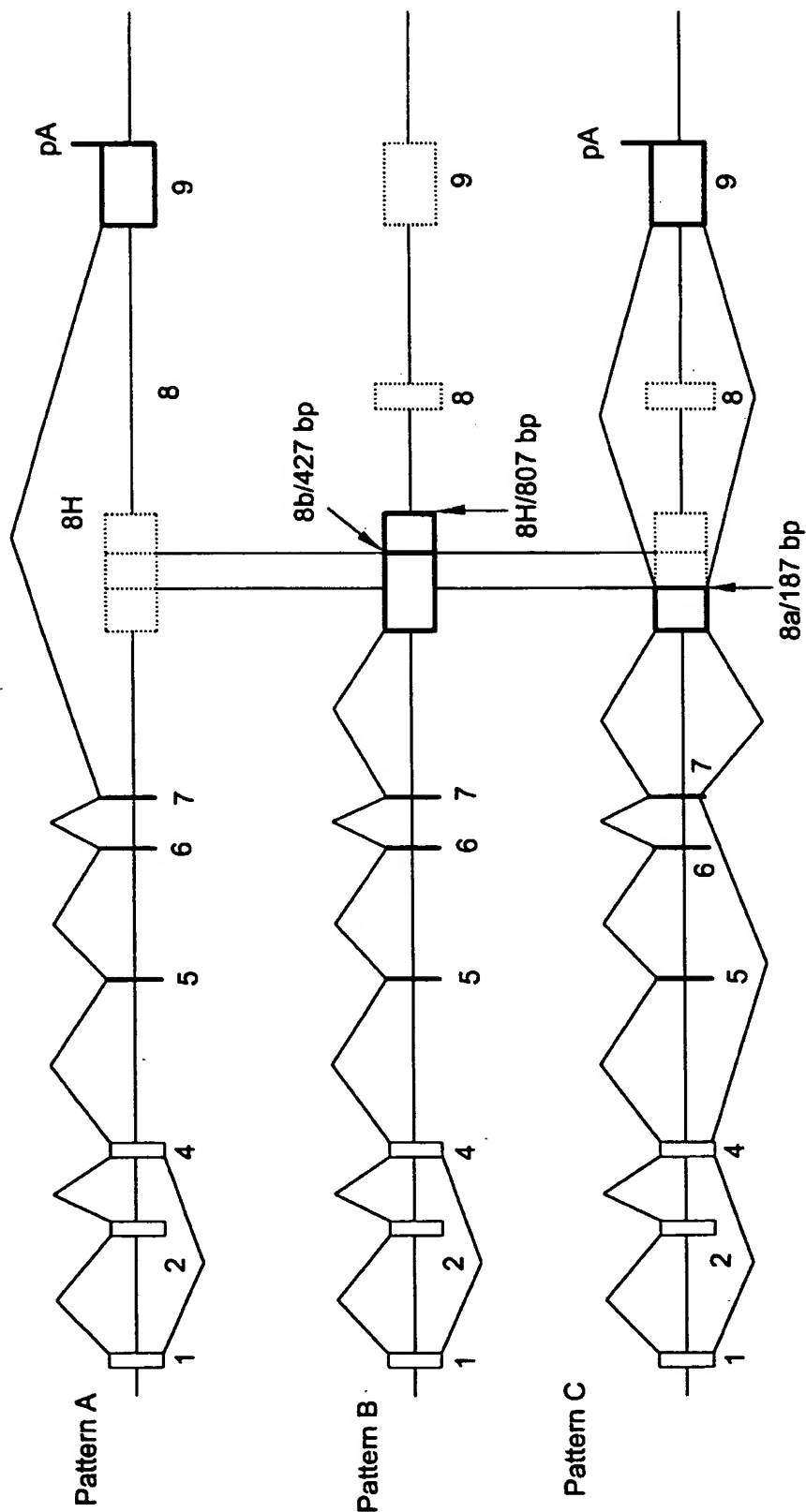


FIG. 3

## SEQUENCE LISTING

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Koike, Chihiro

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<211> 820

<212> DNA

<213> Mus musculus

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<211> 501

<212> DNA

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<210> 7

<211> 3976

<212> DNA

<213> Sus scrofa

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<221> misc\_feature

<222> (580)..(580)

<223> "n" is a gap of from about 600 to about 800 nucleotides

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<221> promoter

<222> (1863)..(2992)

<223> fragments and derivatives of this region have promoter activity.

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<221> 5'UTR

<222> (2463)..(3016)

<223> Untranslated exon 1 runs from about nucleotide 2436 to about nucleotide 3016

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<221> Intron

<222> (1)..(2462)

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gccaagctcc cccaaaactc tgttggcatt tgtcttgagt ttataggttg atgcatggag	360
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<222> (4732)..(4814)

<223> untranslated exon 1A found in some transcripts

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&lt;221&gt; misc\_feature

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&lt;223&gt; untranslated exon 2 found in some transcripts

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (5354)..(8989)

&lt;400&gt; 8

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21

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&lt;210&gt; 9

&lt;211&gt; 240

&lt;212&gt; DNA

&lt;213&gt; Sus scrofa

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (1)..(29)

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (30)..(118)

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (119)..(240)

&lt;220&gt;

&lt;221&gt; 5'UTR

&lt;222&gt; (30)..(38)

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (39)..(41)

&lt;223&gt; This "atg" is the translation start codon

&lt;400&gt; 9

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aattatgaaa catgatgaaa tgatgttgat gaaagtctcc tctaattctcc tagttatcag 180

ccaagtcacc agcttgcat taaaagtagga ttcactgaca ccgtaaagaa agcattccag 240

&lt;210&gt; 10

&lt;211&gt; 2685

&lt;212&gt; DNA

&lt;213&gt; Sus scrofa

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (1)..(2140)

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (2141)..(2176)

&lt;223&gt; This region defines exon 5

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (2177)..(2685)

&lt;400&gt; 10

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tgctaggaag cttagagcaa agtattgtgc ttaaatgctt gcattttcct tggccttcat 120

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tcacagaggc ggctcagatt ccaagttgct gtggctgtgg cgtaggccgg cagctacagc 660

tccaattaga cccctagcct gggaacttcc acatgccgca gggtgcaacc ccaaaagata 720

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ctgttaggca cactcttgct aatccctctt cactgggcct cctatgtatc cttcagaact 840

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25

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<210> 11  
<211> 180  
<212> DNA  
<213> Sus scrofa

<220>  
<221> Intron  
<222> (1)..(37)

<220>  
<221> misc\_feature  
<222> (38)..(100)  
<223> This region defines exon 6

<220>  
<221> Intron  
<222> (101)..(180)

<400> 11  
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tctgtgtatc tgctcaaggc tgtagagtcc aaataaaatg gtttcacagc catgaccttc 180

<210> 12  
<211> 242



<212> DNA

<213> Sus scrofa

<220>

<221> Intron

<222> (1)..(100)

<220>

<221> misc\_feature

<222> (101)..(205)

<223> This region defines exon 7

<220>

<221> Intron

<222> (206)..(242)

<400> 12

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cctccctgcc ttctcagct tgttttccgt ttgtacgtag gactcacagt taccacgaag 120

aagaagacgc tataggcaac gaaaaggaac aaagaaaaga agacaacaga ggagagcttc 180

cgctagtgga ctggtttaat cctgagtaag aaaagaagcg ttgccctatt tcagtaaadc 240

ca 242

<210> 13

<211> 720

<212> DNA

<213> Sus scrofa

<220>

<221> Intron

<222> (1)..(257)

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (258)..(394)

&lt;223&gt; This region defines exon 8

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (395)..(720)

&lt;400&gt; 13

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cagggcagcc cgcagaggca cttgggccag agcctcctgt ccttccccca gaagatgccg 120

caatgtcaca ccaccagctg actggggcta aaatacagtc aggattcaag gccagtccca 180

caagccatga ctgacccatg ttccccaga ctgtcgtacc ttagcaaagc catcctgact 240

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cctcaggggtg aggggcttgg gttgcagga agaaaatctg ctataccac tgcacccaaa 660

gtcgacagta cacccacagc cacctccacc ctgacctcca cggccctctg tggaaattcc 720

&lt;210&gt; 14

&lt;211&gt; 2964

<212> DNA

<213> Sus scrofa

<220>

<221> Intron

<222> (1) .. (318)

<220>

<221> misc\_feature

<222> (319) .. (2904)

<223> This region defines exon 9

<220>

<221> terminator

<222> (1010) .. (1012)

<223> This is the translation termination signal

<220>

<221> 3'UTR

<222> (1012) .. (2964)

<220>

<221> terminator

<222> (2858) .. (2863)

<223> This is one transcription termination signal

<220>

<221> terminator

<222> (2883) .. (2888)

<223> This is one transcription termination signal

<220>

<221> polyA\_signal

<222> (2904) .. (2904)

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cattcttctt tatttcagat acattgagca ttacttggag gagttcttaa tatctgcaaa	360
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<210> 15

<211> 1500

<212> DNA

<213> Sus scrofa

<220>

<221> misc\_feature

<222> (1)..(1500)

<223> genomic sequence between exons 2 and 4

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<210> 16

<211> 500

<212> DNA

<213> Sus scrofa

<220>

<221> misc\_feature

<222> (1)..(500)

<223> genomic sequence about 4-5 kbp downstream from porcine exon 4.

<400> 16

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cagcaacgca ggatctgagc cacgtctgca accttcacca cagctcacgg caacgccaga 300  
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34

ttcgttaagc actgcgccac gacgggaact ccctcattta gaaatattta ttgagcacct 420

actgtatgcc aggcattggt ctaggttcat accaaagaag gctcaaaaag atggcatccg 480

aactgttgcc cttgaaagga 500

<210> 17

<211> 1520

<212> DNA

<213> Mus musculus

<220>

<221> Intron

<222> (1) .. (1130)

<220>

<221> promoter

<222> (381) .. (1321)

<223> Fragments and derivatives have promoter activity.

<220>

<221> 5'UTR

<222> (1131) .. (1320)

<223> untranslated exon 1

<220>

<221> Intron

<222> (1321) .. (1520)

<400> 17

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gtggggcgagc gcgagcgccg ctattccggc ccagccctac ctcggtcctt gcttttgtcc 300

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acagtgtcgc tgcatctgtt tgcttactgg ggtctccgc caccttcctt cgctatccga 420

atagctgata ttcagggcag cacagggcag ggcagggcag ggcagggcga gtagggcaga 480

tcagatcctg ggaccaccgg tactaaccag tgagtgtaga aagcaggagg tgtcttttcc 540

tactgtagtt aggacagggc gggttggtc ttcttatgga caagatggaa aaggggtgca 600

ggtaggggca aagtgagaga cactcgaatt tgagagacag acagactcct aacagtgaag 660

gaaggaccaa gccaaaatca agcctgggca aagtctcagg cactaacttt gctgtgttgg 720

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tatgcgattg cacaaactct tgaaaatcat ccaagaaaca gcaaagcggg aaataatgtt 960

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<210> 18  
 <211> 1207  
 <212> DNA  
 <213> Mus musculus

<220>  
 <221> Intron  
 <222> (1)..(653)

<220>  
 <221> 5'UTR  
 <222> (654)..(773)  
 <223> untranslated exon 2

<220>  
 <221> Intron  
 <222> (774)..(1207)

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 ggtatgggtt tctgaggctg tcccaagtgc atatggtaaa ggcttctcta tggagattta 180  
 caccattttc taaagtgcag tgttccacat aactgtgtgg cttccagagc caggctgtgg 240

37

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atcgcttggt actcctggca tggtttggtc ctgcagtttt tcctctgggt gaggaagtca 480

gaggaccaac ccagagccct gattctgcct tgctgcgtag acctgaatca acagccctga 540

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tctgccacc ccaccctcc tcttctgaa acagctgttt attattttga caggagttag 660

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agaagctcgg ttgctttgct gtttgctttg gagggaacac agctgacgat gaggtatggt 780

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tgttgctcct tcccttttgt ttctgaaaca ggggctgggt gaatgctggc tggggacttt 960

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tctctgtgtg gggtttgggt tggcggggct gagtcttggg cagggcgcgg tgggaggggt 1080

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gtatcacttg agattgattg cattccacat gacactgctc ccagggacag cccggcactc 1200

nnnnnnn 1207

&lt;210&gt; 19

&lt;211&gt; 900

&lt;212&gt; DNA

<213> Mus musculus

<220>

<221> Intron

<222> (1)..(336)

<220>

<221> misc\_feature

<222> (337)..(517)

<223> untranslated exon 3

<220>

<221> Intron

<222> (518)..(900)

<400> 19

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ccatctctgc cctctgtagt actctaggct ctagttgact ccactccact gctgctgctg 180

ttcttggtga tcctcctatg gtactggcaa gtaggtgaaa gaagaagagt gaatattcct 240

tcaccaaatg tccttatgta ggctccagc agaagggtg gctcagatta aagggtgcta 300

cccccatgcc tggatctaaa acttgctttg ttccaggctg actttgaact caagagatct 360

gcttaccoca gtctcctgga attaaaggcc tgtactacat ttgcctggac ctaagatttt 420

catgatcact atgcttcaag atctccatgt caacaagatc tccatgtcaa gatccaagtc 480

agaaacaagt cttccatcct caagatctgg atcacagggtg tgcccttctg tttctggatt 540

atagttcatc ccagatgtag tcaagttgac cactaggaat agccatcaca agcccgttgt 600

ggaggctgcc cctgcccc cgccccgcgc gccctgagg ctctcaccce tttcttggtg 660

cagctcttgt cttcatctcc agtgtacaac tgtcattccc actctgcac ttgccttcct 720  
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aatcttatgc agaatttaaa aaatacctga ctccttcagt agttccagtt gtttgctggc 840  
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<210> 20

<211> 1020

<212> DNA

<213> Mus musculus

<220>

<221> Intron

<222> (1)..(479)

<220>

<221> misc\_feature

<222> (480)..(568)

<220>

<221> 5'UTR

<222> (480)..(489)

<220>

<221> misc\_feature

<222> (490)..(492)

<223> This "atg" is the translation initiation codon

<220>

<221> Intron

<222> (569)..(1020)

<400> 20

40

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gacacttgct gctgagtga gttacaaatgt atcctgttag gaaggatgtg ggcagatgcc 180

tttcattatc ttactgcat caaacatctt atgggtatga gtgttttgcc tgcaagtatg 240

tatatgtacc acttgtatat gtggacccca tggaggccag aagagcatca ggtcctgtga 300

aaccagagtt atggacacct gtgagctgca aatgtggatg ctgggaactg aatcgagcag 360

gtgtttcatt gaggtgtttc aaccacacag ctgtttctcc agccccagaa gccatctctc 420

attccagatt tagtttatct aatctatttc cccctctttt tttctccctg cctctacagg 480

agaaaataat gaatgtcaag ggaaaagtaa tcctgttgat gctgattgtc tcaaccgtgg 540

ttgtcgtggt ttgggaatat gtcaacaggt aattatgaag ccagctagaa aggctgcttt 600

cattccctgt gactggtgcc agctgagtga ccaatcagtc tgaacataag ggacggagcc 660

gtgagcagga gtccagtctt cctgtgttcc tgagccccag atggccatta aaactgtaga 720

ccatccaagt cacttctgcc ttagtaatta tcctctttca tgccgtgctc ctcaaaccctc 780

gaatttctgt aagctagatg gagagagaaa gtacattaag ccaaaccac catctcaagt 840

aatttgata agcagatccc agaagattca ggccaggcag ggtagtgcac gtatggagtc 900

cttgtgcttg caaggcagag gcaggagcat catacaaatg gaagaccaag cttgtcttca 960

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&lt;210&gt; 21

&lt;211&gt; 1020

&lt;212&gt; DNA

<213> Mus musculus

<220>

<221> Intron

<222> (1)..(584)

<220>

<221> misc\_feature

<222> (585)..(620)

<223> exon 5

<220>

<221> Intron

<222> (621)..(1020)

<400> 21

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tcacttgatt tttttttttt agcccctaaa gttgatttcc tctcttcaag ccagccaatg 720  
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<210> 22

<211> 1020

<212> DNA

<213> Mus musculus

<220>

<221> Intron

<222> (1)..(595)

<220>

<221> misc\_feature

<222> (596)..(661)

<223> exon 6

<220>

<221> Intron

<222> (662)..(1020)

<400> 22

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43

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&lt;210&gt; 23

&lt;211&gt; 912

&lt;212&gt; DNA

&lt;213&gt; Mus musculus

&lt;220&gt;

&lt;221&gt; Intron

<222> (1)..(389)

<220>

<221> misc\_feature

<222> (390)..(491)

<223> exon 7

<220>

<221> Intron

<222> (492)..(912)

<400> 23

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<211> 608  
<212> DNA  
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<223> exon 8

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<221> Intron  
<222> (360)..(608)

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46

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<223> exon 9

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<222> (1088)..(1090)

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<222> (1091)..(3010)

<220>  
<221> Intron

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&lt;400&gt; 25

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&lt;213&gt; Mus musculus

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gaaaataatg aatgtcaaag gaaaagtaat tctgtcaatg ctggttgtct caactgtgat 180  
  
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62

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&lt;210&gt; 31

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&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 31

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<222> (631)..(666)  
<223> This is exon 5

<220>

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<222> (667)..(732)

<223> This is exon 6

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<223> This is exon 8h

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66

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&lt;210&gt; 33

&lt;211&gt; 3322

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 33

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<212> DNA

<213> Homo sapiens

<400> 34

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73

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&lt;210&gt; 35

&lt;211&gt; 244

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

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&lt;223&gt; 5' flanking sequence

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<222> (61)..(149)

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<220>  
<221> Intron  
<222> (1)..(60)

<220>

<221> misc\_feature

<222> (61)..(176)

<223> human untranslated exon 7

<220>

<221> Intron

<222> (177)..(242)

<400> 39

tctttgacca ccgcaatcac cttccctgcc ttacctggtt tactttccct ttgtacttag 60

gatccacaat tatcaacaag gggaagaaga catagacaaa gaaaaaggaa gagaggagac 120

caaaggaagg aaaatgacac aacagagctt cggctatggg actggtttaa tccaaagtaa 180

gaaaagcggc gtcactccct gtgcagcaaa tccatggccc tgcagggggt ggtgtggcnn 240

nn 242

<210> 40

<211> 487

<212> DNA

<213> Homo sapiens

<220>

<221> Intron

<222> (1)..(60)

<220>

<221> misc\_feature

<222> (61)..(487)

<223> a version of human untranslated exon 8h

<400> 40

78

atagaatatt ttaattttta attcaacata aatttttaag ggtgctgttt tttcttccag 60  
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acctggactg agagatcatg cggttaagga gtgtgtaaca ggcggaccac ctgttgggac 180  
tgcgagattc tcaaggggaa ggactgggtc tcatttctcc catctcagcg cttagcagga 240  
tgacctggta tagagcaggg aactgggaaa tgtgggtcag gggatcagac actccagttg 300  
ggtcttttat ataaattaaa tggcaaaagg ctccataccc ttctccttct ttcctaccct 360  
ccactttatc tgcaaaatgg gaatgatgat aacacccact tcatagaatg gtcatgaaga 420  
tcaaatgaga gaataaaaagt caagcactta gcctctggtg cacaataagt attaaataag 480  
tatacct 487

&lt;210&gt; 41

&lt;211&gt; 454

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)..(380)

&lt;223&gt; a version of the human untranslated exon 8h

&lt;220&gt;

&lt;221&gt; Intron

&lt;222&gt; (381)..(454)

&lt;400&gt; 41

attcctcctt ttcctttttt aaaaataata ttaccaaag tccagcttat acacatttac 60  
aagacttagc tagtgggcta tgtagagct actaaaagat ctttgacaag ctaaaactaa 120

gatgcaatga atgaggtgta acgaacaaga gagttttaag ttcagaaatg gttacagaag 180  
tataagacag ctgtgtgggt gttttttggt ttttggtttc tggtttacia tctcgtcatt 240  
caacaaagat gggagtttta tagaactaaa agcaccatgt aagctactaa aaacaacaac 300  
aaaaaaggct catcatttct cagtctgaat tgacaaaaat gccaatgcaa ataaaaatga 360  
ttacttttta tttttcaacg ttgtttgttt atttatttat ttcgagatgg agtttcactc 420  
ttgttgccct ggctggagtg cagtggcgcn nnnn 454

<210> 42

<211> 2848

<212> DNA

<213> Homo sapiens

<220>

<221> Intron

<222> (1)..(65)

<220>

<221> misc\_feature

<222> (66)..(2676)

<223> human untranslated exon 9

<220>

<221> misc\_feature

<222> (2677)..(2848)

<223> an inter-gene sequence

<400> 42

ttcagcttgt ggtttctttc aggaatccca gaggataaat gttttgcttt tcttctttgt 60

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gttggccaca aagtcatttt ttacatcatg gtggatgatg tctccaagct gccgtttata	180
gagctgggtc ctctgcattc cttcaaaatg tttgaggta agccagagaa gaggtggcaa	240
gacatcagca tgatgcgtat gaagatcact ggggagcaca tcttggccca catccaacac	300
gaggtcgact tcctcttctg catggatgtg gaccaggtct tccaagacca ttttggggtg	360
gagaccctag gccagtcagt ggctcagcta caggctggcg gtacaaggca gatccctatg	420
actttaccta ggagaggtgg aaagagtcag caggatacat tccatttggc caggggattt	480
ttattacat gcagccattt ctggaggaac acccattcag gttctcaaca tcaccagga	540
gtgctttaag ggaatcctcc tggacaagaa aaatgacata gaagccaagt ggcattgatga	600
aagccaccta aacaagtatt tccttctcaa taaaccctct aaaatcttat ccctaaaata	660
ctgctgggat tatcatatag gcctgccttc agatattaaa actgtcaagt gatcgtggca	720
gacaaaagag tataatttgg ttagaaataa tgtctgactt caaattgtgc cagtagattt	780
ctgaatttaa gagagagaat attctggcta ctctctcaga aaagtaacac ttaattttaa	840
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tggggctttg taatgtggaa gaatgaatct agggcaatca gatataaatt ccagtgatt	960
tcttatctat tctgggtttg ggggaaatac tatcaactga accaaaaata actgtcata	1020
ggcagagata aagccagaaa cactctacac atgccagatg acatctggag aaaagggtgc	1080
taagggaagc gtttggcagc aagatatgat tgtaaggggt tgtcccttga gttcaatgtc	1140
tgcctatttc tgatgggtct aaagcaacat ggagttactg tgcagcagaa ctctcagtaa	1200

agacaccatt tgccttggca atcctcaaaa agcttcaata gcagattgct tcagaccatc 1260

tgtagtccgt ccttttctca tctggatggt gtttggttc tgtgcgaaag attggtggag 1320

tgtcccagta gatatcatgg tgggtgtgta tcagagtccc aaggaacctg aatgagccaa 1380

ggtgcccagc atgaagtcaa aacaaagcct tgacatgagt ttgcatgaa atagcgaaga 1440

gagagtggaa gagaggagcc aatcactgtg gggcagtgcc accctgaggg cacttagggt 1500

atgggggttg tgcttaaata catcacagat ccaggctactg aatgggagga agtgtgggtg 1560

atttccaatc tcattgacct tatgttcagg gacttgaacg gaagatgttt cttgtgttgc 1620

ctaagtggta ttcagtctac cagactctgc aacttgcac ttcaaatacct tggtaaagag 1680

atgtggatgg tgtcagagaa ggcaaaggcc tgcagtggat tgaagaggct tgcaagcagt 1740

tctgtttcta ggatgtgggc ttcacagaa gacactcggc caccacttag ctagtctaaa 1800

cctcagggtt cctcagccca tcatacccca acttgaggga ctgacatcaa ggagtagact 1860

ggagaaacag cctcccatc aagtaacctc ttgttctctc ctgctccatc tgcactatag 1920

aagtgttaata attagacata cttggcaaaa tggctaattg atttggtaac agaagcatga 1980

gccataacaa tggaagatct agttatcatg actgaacagc ttaacattca attcccttct 2040

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gttaggagac agtgtgtacc tatgcacgta tatttatgtt ttgcttgtgt tccagtctcg 2160

gtcatttgtt tccattttca agcaatttat ttgaagagcc attgcactag cctgatgtat 2220

actgcaatga gcttctttga taaaatgaaa cttaaatttt tctcgaccat ttcaccgtgc 2280

ctcctacttc attttttgcc agaaaatctc acatccaaca aaacaaaaca aaaaccctga 2340



attagtgggc tttgaaaagg aaaaagcagg gctttgaaaa agtagatcac acatcagtta 2400  
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ccccaatgga caaattcata atcttggtta tcgttattac taaacttttt aaaaaatgtc 2640  
ccaatttaca attaaataaa ttactttctc agtatattct ggtctggtca tggattgtgc 2700  
atttcctccc aaagatatcc aaaattgtca attagagaat ttaggtttt cagactcaga 2760  
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<210> 43

<211> 2303

<212> DNA

<213> Rhesus monkey

<220>

<221> misc\_feature

<222> (1)..(44)

<223> This is exon 1

<220>

<221> misc\_feature

<222> (45)..(159)

<223> This is exon 2

<220>

<221> misc\_feature

<222> (160)..(278)

<223> This is exon 3

<220>

<221> misc\_feature

<222> (279)..(367)

<223> This is exon 4

<220>

<221> misc\_feature

<222> (368)..(403)

<223> This is exon 5

<220>

<221> misc\_feature

<222> (404)..(469)

<223> This is exon 6

<220>

<221> misc\_feature

<222> (470)..(584)

<223> This is exon 7

<220>

<221> misc\_feature

<222> (585)..(2260)

<223> This is exon 9

<220>

<221> misc\_feature

<222> (543)..(545)

<223> This is an early stop codon

<220>

<221> misc\_feature

<222> (1276)..(1278)

<223> This is a stop codon

<220>

<221> misc\_feature

<222> (1225)..(1260)

<223> This is a polyadenylation signal

<400> 43

gctcgctgcg cgccggctct gggtgccagg gttctgcgga tcaggagttg aaaccagcat 60

cttcccttca tctgagtcct gcctccttct gcagaaggga gctcaaaaga actttgttgt 120

tttgcctttt actctggggt gaaagcaaca gacgataagg atctcactct gtcgccaag 180

ctggagtgca gtggcttgat tacagctcac tgtagcctgg accttccaag gctctgggtg 240

atcttcttac ctgagcttcc ccagtagctg gactacagga gaaaataatg aatgtcaaag 300

gaaaagtaat tctgtcaatg ctggttgtct caactgtgat cattgtgttt tgggaatata 360

tcaatagccc agaaggttct ttcttgggga tgtatcgctc aaaaaacca gaggttgatg 420

acagcagtgc tcagaagagc tggtggtttc cgagctggtt taacaatggg atccacaatc 480

atcaacaaga ggaagaagac atagacaaaa aagaggaaga gaggagacca aagaaaggaa 540

gatgacacaa cagagcttcg gctatgggac tgatttaatc caaaatatat tgagcattac 600

ttggaagagt tcataacacc tgctaataagg tacttcaagg tcggccacaa agtcatattt 660

tacattatag tggatgatgt ctccaagggtg ctgtttatag agctgggtcc tctgcattcc 720

ttaaaagtgt ttgaggtcaa gccagagaag aggtggcaac acatcagcat gatgcctgtg 780

aagatcatca gggagcacat cttggccac atccaacacg aggtcgactt cctcttctgc 840

atggatgtag accaggtcctt ccaagacaat tttggggtga agaccctagg tcagtcagtg 900

gctcagctac agccctggtg gtacaaggca gatcctgatg actttaccta ggagaggcag 960

aaagagtcag cagcatgcat tccatttggc caggaggatt tttattacca cacagccatt 1020

tttggaggaa cacccattca ggttctcaac atcccccagg agtgctttaa gagaatcctc 1080

ctggaaaaga aaaatgacat agaagctgag tggcatgatg aaagccacct aaaccagtat 1140

ttccttctca acaaaccctc taaaatctta tccctagaat actgctggga ttatcatatc 1200

agcctgcctt cagatattaa aactgtcaag cggtcgtggc agacaaaaga gtataatttg 1260

gtagaaaata tcctctgact tcaaattgtg ccagtagatt tctgaatttg agagaggagt 1320

attctggctg cttcctcaga aaagtaacac ttaattttta gttaaaaaaa atactaatga 1380

aacaccaaca tggcaaacac ataccattcc ttcttgtaac ttgaggcttt gtaatgtggg 1440

agaatgaatc tagggtaatc agatgtaaat tccagtgat ttcttatcta ttttgggttt 1500

gggggaaata ctatcaactg aacaaaaaag aacttgtcat aggcaaagat aaagccagaa 1560

acactctaca catgccacat aacatctgga gaaaagggtg ctaagggaag cgtttggcag 1620

caagatatga ttgtaagggg ttgtcccttg agttcaatgc ctgcctatctt ccaatggatc 1680

taaaacaacg tgaagttact gtgcagcaga gctctcagta aggacacat ttgccttggc 1740

aatcctcaaa attcttcaat agcagattgt ttcaggccat ctgtagtctg tccttttctc 1800

atcaggatgt tgtttggctt ctgtgcgaaa aattgggtgga gtgtcctggt agatattgaa 1860

actaggcctc atatagaaaa aattaacacc aggtggctct ggatagagtc cgcacctgcc 1920  
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gccatagcta gacccaatca ttttgcgct taagctttgt ttgaatttcg cgccctaagc 2160  
tgtgtttgaa cttgtgtttg cctatataaa cagcctgtaa caagcagtcg ggggtcccagg 2220  
gccaaacttag agcttgggac cctagcgcg ctagtaataaa taactctctg ctgcgaaaaa 2280  
aaaaaaaaa aaaaaaaaaa aaa 2303

<210> 44

<211> 2630

<212> DNA

<213> Rhesus monkey

<220>

<221> misc\_feature

<222> (1)..(44)

<223> This is exon 1

<220>

<221> misc\_feature

<222> (45)..(159)

<223> This is exon 2

<220>

<221> misc\_feature

<222> (160)..(278)

<223> This is exon 3

<220>

<221> misc\_feature

<222> (279)..(367)

<223> This is exon 4

<220>

<221> misc\_feature

<222> (368)..(403)

<223> This is exon 5

<220>

<221> misc\_feature

<222> (404)..(469)

<223> This is exon 6

<220>

<221> misc\_feature

<222> (470)..(584)

<223> This is exon 7

<220>

<221> misc\_feature

<222> (585)..(911)

<223> This is exon 8

<220>

<221> misc\_feature

<222> (912)..(2587)

<223> This is exon 9

<220>  
 <221> misc\_feature  
 <222> (288)..(290)  
 <223> This is a putative start codon

<220>  
 <221> misc\_feature  
 <222> (543)..(545)  
 <223> This is a putative stop codon

<220>  
 <221> misc\_feature  
 <222> (1603)..(1605)  
 <223> This is a putative stop codon

<220>  
 <221> misc\_feature  
 <222> (2582)..(2587)  
 <223> polyadenylation signal

<400> 44  
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 ttgacctttt actctggggt gaaagcaaca gacgataagg atctcactct gtcgccaag 180  
 ctggagtgca gtggcttgat tacagctcac tgtagcctgg accttccaag gctctgggtg 240  
 atcttcctac ctgagcttcc ccagtagctg gactacagga gaaaataatg aatgtcaaag 300  
 gaaaagtaat tctgtcaatg ctgggtgtct caactgtgat cattgtgttt tgggaatata 360

tcaatagccc agaaggttct ttcttgggga tgtatcgctc aaaaaaccca gaggttgatg	420
acagcagtgc tcagaagagc tggtaggttc cgagctggtt taacaatggg atccacaatc	480
atcaacaaga ggaagaagac atagacaaaa aagaggaaga gaggagacca aagaaaggaa	540
gatgacacaa cagagcttcg gctatgggac tgatttaatc caaagaaacg cccagagggtg	600
gtgagagtga ccagatggaa ggcaccgggt gtgtggaaag gcacttaca caaagccatc	660
ctaggaaatt attatgccaa acagaaaatt acggtgggat tgaaggcttt tgctattgga	720
agtgggtgtc actgatgaaa ctgtccttga ctatttcttg ttccactgtc aagacathtt	780
tgtggagact cctgaactga tggaggccag ccatgatttt ttgatttatt agatagaaga	840
atgttttcat ggaactgttt tagtctcctt tctgtgagg ccctaaaatg ctgagaacaa	900
aataagagta gatatttga gcattacttg gaagagtcca taacacctgc taataggtac	960
ttcaagggtcg gccacaaagt catattttac attatagtgg atgatgtctc caagggtgtg	1020
tttatagagc tgggtcctct gcattcctta aaagtgtttg aggtcaagcc agagaagagg	1080
tggcaacaca tcagcatgat gcctgtgaag atcatcaggg agcacatctt ggcccacatc	1140
caacacgagg tcgacttcct cttctgcatg gatgtagacc aggtcttcca agacaatttt	1200
ggggtgaaga ccctaggtca gtgagtggt cagctacagc cctgggtgga caaggcagat	1260
cctgatgact ttacctagga gaggcagaaa gagtcagcag catgcattcc atttgccag	1320
gaggattttt attaccacac agccattttt ggaggaacac ccattcaggt tctcaacatc	1380
ccccaggagt gctttaagag aatcctcctg gaaagaaaa atgacataga agctgagtgg	1440
catgatgaaa gccacctaaa ccagtatttc cttctcaaca aaccctctaa aatcttatcc	1500



ctagaatact gctgggatta tcatatcagc ctgccttcag atattaaaac tgtcaagcgg 1560

tcgtggcaga caaaagagta taatttggtt agaaatatca tctgacttca aattgtgcc 1620

gtagatttct gaatttgaga gaggagtatt ctggctgctt cctcagaaaa gtaacactta 1680

attttaagtt aaaaaaata ctaatgaaac accaacatgg caaacacata ccattccttc 1740

ttgtaacttg aggctttgta atgtgggaga atgaatctag ggtaatcaga tgtaaattcc 1800

cagtgatttc ttatctattt tgggtttggg ggaaatacta tcaactgaac caaaaagaac 1860

ttgtcatagg caaagataaa gccagaaaca ctctacacat gccacataac atctggagaa 1920

aagggtgcta agggaagcgt ttggcagcaa gatatgattg taaggggttg tcccttgagt 1980

tcaatgcctg cctattttcca atggatctaa aacaacgtga agttactgtg cagcagagct 2040

ctcagtaagg acaccatttg ccttggcaat cctcaaaatt cttcaatagc agattgtttc 2100

aggccatctg tagtctgtcc ttttctcatc aggatgttgt ttggcttctg tgcgaaaaat 2160

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ccaattagca taagacgcct tgctcaggcc atagctagac ccaatcattt tgcgcttaa 2460

gctttgtttg aatttcgcg cctaagctgt gtttgaactt gtgtttgcct atataaacag 2520

cctgtaacaa gcagtcgggg tcccagggcc aacttagagc ttgggaccct agcgcgctag 2580

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<210> 45  
<211> 35  
<212> DNA  
<213> Artificial/Unknown

<220>  
<221> misc\_feature  
<222> ()..()  
<223> Antisense primer for cloning porcine exon 4

<400> 45  
ctgttgatgt attcccaaaa cacaaccatt acagt

35

<210> 46  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> Antisense primer for cloning porcine exon 4

<400> 46  
agacaagcag cattgacaga accactc

27

<210> 47  
<211> 25  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature

<222> ()..()

<223> Antisense primer for cloning porcine exon 2

<400> 47

ctcatcctct gcttctctcc cccca

25

<210> 48

<211> 26

<212> DNA

<213> artificial sequence

<400> 48

ccccccagag taaaaggcga aacaag

26

<210> 49

<211> 25

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> Sense primer for cloning porcine exon 2

<400> 49

aacgcagcac cttcccttcc tccca

25

<210> 50

<211> 25

<212> DNA

<213> artificial sequence

<400> 50

cttgtttcgc cttttactct ggggg

25

<210> 51

<211> 23

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> Sense primer for cloning porcine exon 1

<400> 51

gccactgttc' cctcagccga gga

23

<210> 52

<211> 24

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> Sense primer for cloning porcine exon 1

<400> 52

cgagcgcacc cagcttctgc cgat

24

<210> 53

<211> 24

<212> DNA

<213> artificial sequence

94

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; Antisense primer for cloning porcine exon 1

&lt;400&gt; 53

tgcgctcggg gatggccctc tcct

24

&lt;210&gt; 54

&lt;211&gt; 24

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; Antisense primer for cloning porcine exon 1

&lt;400&gt; 54

ggcgtcctcg gctgaggga cagt

24

&lt;210&gt; 55

&lt;211&gt; 28

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; Sense primer for cloning porcine exon 1A

&lt;400&gt; 55

cagaacaact tctgaagcct aaaggatg

28

<210> 56

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> Sense primer for cloning porcine exon 1

<400> 56

caaatggtgg atcggacctc ccaggct

27

<210> 57

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> Sense primer for cloning porcine exon 1

<400> 57

agtactgggt gatagacccc actccac

27

<210> 58

<211> 25

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature  
<222> ()..()  
<223> Sense primer for cloning porcine exon 1

<400> 58  
gcgcagggct ccggggcccc tccct 25

<210> 59  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> Sense primer for cloning porcine exon 9

<400> 59  
ctgggattat catataggca tgtctgt 27

<210> 60  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> Sense primer for cloning porcine exon 9

<400> 60  
agagtattac tctggctact tctccag 27

<210> 61  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for identifying 5' flanking region of murine exon 1

<400> 61  
ctgagagcgc gaggtcttca gcagaat 27

<210> 62  
<211> 28  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for identifying 5' flanking region of murine exon 1

<400> 62  
cttctcattc caagaagagt cttacaag 28

<210> 63  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature



<222> ()..()

<223> primer for identifying 3' flanking region of murine exon 1

<400> 63

cctgcctttt cttagctggc tgacacc

27

<210> 64

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying 3' flanking region of murine exon 1

<400> 64

cttgtagact cttcttgga tgagaag

27

<210> 65

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying 5' flanking region of murine exon 2

<400> 65

catcgtcagc tgtgttcct ccaaagc

27

99

&lt;210&gt; 66

&lt;211&gt; 27

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying 5' flanking region of murine exon 2

&lt;400&gt; 66

aaagcaaccg agcttctgtc gagctct

27

&lt;210&gt; 67

&lt;211&gt; 38

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying murine exons 2 and 3

&lt;400&gt; 67

gtaccttctt ttctctgtct gagccctgcc tccttcgg

38

&lt;210&gt; 68

&lt;211&gt; 35

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

100

<223> primer for identifying murine exons 2 and 3

<400> 68

agatcttgag gatccaagac ttgtttctga cttgg

35

<210> 69

<211> 34

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exons 3 and 4

<400> 69

gctgactttg aactcaagag atctgcttta cccc

34

<210> 70

<211> 28

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exons 3 and 4

<400> 70

ctgttgacat attcccaaaa cacgacaa

28

<210> 71

101

<211> 30  
<212> DNA  
<213> artificial sequence  
  
<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for identifying murine exons 4 and 5

<400> 71  
gtcaaggga aagtaatcct gttgatgctg

30

<210> 72  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for identifying murine exons 4 and 5

<400> 72  
tatccacaag aaagagccgt ctgggct

27

<210> 73  
<211> 27  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for identifying murine exons 5 and 6

<400> 73

agcccagacg gctctttctt gtggata

27

<210> 74

<211> 34

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exons 5 and 6

<400> 74

ccagcttggg aaccaccagt ccttctgcc a tctg

34

<210> 75

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exons 6 and 7

<400> 75

ttccagaggt tggtgagaac agatggc

27

<210> 76

<211> 33

103

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying murine exons 6 and 7

&lt;400&gt; 76

gcgatctcca tttctaccct tttctctccg tcc

33

&lt;210&gt; 77

&lt;211&gt; 28

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying murine exon 7

&lt;400&gt; 77

caagaagaca acgtagaagg acggagag

28

&lt;210&gt; 78

&lt;211&gt; 27

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying murine exon 7

<400> 78

tcgcattgaa gagcctcagc tatggga

27

<210> 79

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying exon 8

<400> 79

ccacagtgag tttctgtgtg gcgatgt

27

<210> 80

<211> 28

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exon 8

<400> 80

agagctgtgt cataagtgcc ttcccaca

28

<210> 81

<211> 27

<212> DNA

105

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exon 8

<400> 81

gatgttttga cagtgacccc gtggaag

27

<210> 82

<211> 28

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exon 8

<400> 82

tgtgggaagg cacttatgac acagctct

28

<210> 83

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying murine exon 9



106

&lt;400&gt; 83

agaggggttca ggtgcacgac aggcac

27

&lt;210&gt; 84

&lt;211&gt; 27

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying murine exon 8

&lt;400&gt; 84

gtacatgtca gcagactcca gaaagtc

27

&lt;210&gt; 85

&lt;211&gt; 27

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for identifying 3' flanking region of murine exon 9

&lt;400&gt; 85

gactttctgg agtctgctga catgtac

27

&lt;210&gt; 86

&lt;211&gt; 27

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying 3' flanking region of murine exon 9

<400> 86

gatgcctgtc gtgcacctga accctct

27

<210> 87

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying 3' flanking region of murine exon 9

<400> 87

aggccattgc accatcttgg tgaacag

27

<210> 88

<211> 28

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for identifying 3' flanking region of murine exon 9

<400> 88

gatcttacct ttgtccacag ggctctac

28

<210> 89

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for obtaining murine promoter

<400> 89

ccaatgcatc ttttcccagt gggctct

27

<210> 90

<211> 27

<212> DNA

<213> artificial sequence

<220>

<221> misc\_feature

<222> ()..()

<223> primer for isolation of transcription initiation site

<400> 90

cccagaacag atctgactgc ctctttc

27

<210> 91

<211> 27

<212> DNA

<213> artificial sequence

109

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for isolation of transcription initiation site

&lt;400&gt; 91

agttttgctt gtctgggcca ctatcgg

27

&lt;210&gt; 92

&lt;211&gt; 27

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for isolation of transcription initiation site

&lt;400&gt; 92

gactggagag agtgctgtcc tccttgc

27

&lt;210&gt; 93

&lt;211&gt; 29

&lt;212&gt; DNA

&lt;213&gt; artificial sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for cloning Rhesus alpha 1,3 GT

&lt;400&gt; 93

gaggtcaagc cagagaagag gtggcaaca

29

<210> 94  
<211> 30  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for cloning Rhesus alpha 1,3 GT

<400> 94  
gacttcctct tctgcatgga tgtagaccag

30

<210> 95  
<211> 29  
<212> DNA  
<213> artificial sequence

<220>  
<221> misc\_feature  
<222> ()..()  
<223> primer for cloning Rhesus alpha 1,3 GT

<400> 95  
atgtcgagaa cctgaatggg tggtcctcc

29

<210> 96  
<211> 30  
<212> DNA  
<213> artificial sequence

<220>

111

&lt;221&gt; misc\_feature

&lt;222&gt; ()..()

&lt;223&gt; primer for cloning Rhesus alpha 1,3 GT

&lt;400&gt; 96

ctggccaaat ggaatgcatg ctgctgactc

30

(19) World Intellectual Property Organization  
International Bureau



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3 May 2001 (03.05.2001)

PCT

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- (72) Inventor; and
- (75) Inventor/Applicant (for US only): KOIKE, Chihiro [US/US]; 5628 Hempstead Street, Pittsburgh, PA 15206-1520 (US).
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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).
- Published:  
— with international search report
- (88) Date of publication of the international search report:  
31 January 2002
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 01/30992 A3

(54) Title:  $\alpha$ 1-3 GALACTOSYLTRANSFERASE GENE AND PROMOTER

(57) Abstract: The present invention provides a recombinant expression cassette comprising an  $\alpha$  1-3 galactosyltransferase promoter operably linked to a polynucleotide for expression. The invention also provides a recombinant mutating cassette comprising a region of homology to an  $\alpha$  1-3 galactosyltransferase genomic sequence. The cassettes can be employed to express foreign genes or to disrupt the native  $\alpha$  1-3 galactosyltransferase genomic sequence, particularly within an animal. Thus, the invention also provides transgenic animals and methods for their production and use.

# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/29139

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/54 C12N5/10 C12N15/85 C12N9/10 A01K67/027

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N A01K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, BIOSIS, EMBL

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 849 991 A (CRAWFORD ROBERT J ET AL) 15 December 1998 (1998-12-15) the whole document	13,14, 17-43
X	WO 95 28412 A (GUSTAFSSON KENTH T ;INST OF CHILD HEALTH (GB); BAETSCHER MANFRED W) 26 October 1995 (1995-10-26) claims 1-13 example 6	13,14, 17-43
A	KATAYAMA A. ET AL.: "Porcine a-1,3-galactosyltransferase: full length cDNA cloning, genomic organization and analysis of splicing variants." GLYCOCONJ. J., vol. 15, 1998, pages 583-589, XP002159948 the whole document	1-43

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

9 February 2001

Date of mailing of the international search report

29. 05. 2001

Name and mailing address of the ISA

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Authorized officer

Galli, I



# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US 00/29139

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
  
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
  
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
  
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
  
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

see further information sheet invention 1.

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-12 and partly 13-43

A recombinant expression cassette comprising an alpha-1,3-galactosyltransferase promoter operably linked to a polynucleotide for expression, other than a polynucleotide encoding alpha-1,3-galactosyltransferase.

A recombinant mutating cassette comprising a first region of homology to an alpha-1,3-galactosyltransferase genomic sequence adjacent to either a second region of homology to said alpha-1,3-galactosyltransferase genomic sequence or a polynucleotide for insertion, WHEREIN a region of homology is homologous to a promoter of said A alpha-1,3-galactosyltransferase gene.

Corresponding vectors, recombinant chromosomes, transgenic cells, embryos, organs and animals.

2. Claims: 13-43, partly

As far as not covered by invention 1:

A recombinant mutating cassette comprising a first region of homology to an alpha-1,3-galactosyltransferase genomic sequence adjacent to either a second region of homology to said alpha-1,3-galactosyltransferase genomic sequence or a polynucleotide for insertion.

Said recombinant cassette, wherein the alpha-1,3-galactosyltransferase sequence is from pig.

Corresponding vectors, recombinant chromosomes, transgenic cells, embryos, organs and animals.

3. Claims: 13-43, partly

Idem as subject matter 2, but wherein the alpha-1,3-galactosyltransferase sequence is from mouse.

4. Claims: 13-43, partly

Idem as subject matter 2, but wherein the alpha-1,3-galactosyltransferase sequence is from man.

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 00/29139

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 5849991 A	15-12-1998	AU 695373 B	13-08-1998
		BR 9506652 A	02-09-1997
		CA 2181433 A	03-08-1995
		EP 0755451 A	29-01-1997
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		AU 711144 B	07-10-1999
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		WO 9520661 A	03-08-1995
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		EP 0755402 A	29-01-1997
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